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(54) Title: SUGARCANE PLANT PROMOTERS TO EXPRESS HETEROLOGOUS NUCLEIC ACIDS

(57) Abstract: Isolated nucleic acids corresponding to sugarcane plant gene promoters are provided. The promoters are capable of directing expression in many or all cells of a plant, preferentially in stem tissue or preferentially in meristem tissue. The isolated promoters are used in the production of transgenic plants, preferably transgenic sugarcane, to direct expression of heterologous nucleic acids in either a constitutive or tissue-specific manner.

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TITLE

Sugarcane plant promoters to express heterologous nucleic acids.

FIELD OF THE INVENTION

THIS INVENTION relates generally to isolated nucleic acid
5 promoters for use in plant genetic engineering. More particularly, the present
invention relates to constitutive and tissue-specific promoters for expression of
heterologous nucleic acids, such as foreign or endogenous coding sequences, in
monocotyledonous plants. The invention is also concerned with a chimeric nucleic
acid construct comprising a promoter of the invention operably linked to the
10 heterologous nucleic acid, expression vectors, transformed plants, cells and tissues
comprising an isolated promoter of the invention.

BACKGROUND OF THE INVENTION

A primary goal of genetic engineering is to obtain plants having
improved characteristics or traits. Many different types of characteristics or traits are
15 considered advantageous, but those of particular importance include resistance to
plant diseases, resistance to viruses or insects and resistance to herbicides. Other
advantageous characteristics or traits include tolerance to cold or soil salinity,
enhanced stability or shelf life of the ultimate consumer product obtained from a
plant, or improvement in the nutritional value of edible portions of a plant.

20 Recent advances in genetic engineering have enabled the
incorporation of a selected gene (or genes) into plant cells to impart a desired quality
(or qualities) to a plant of interest. The selected gene (or genes) may be derived from
a source different from the plant of interest or may be native to the desired plant, but
engineered to have different or improved qualities. This new gene (or genes) may
25 then be expressed in cells of the regenerated plant to exhibit the new trait or
characteristic.

In order for the newly incorporated gene to express the protein for
which it codes in a plant cell, the proper regulatory signals must be present and in the
proper location with respect to the gene. These regulatory signals include a promoter,
30 a 5' non-translated leader sequence and a 3' polyadenylation signal.

Generally, the efficiency of gene expression is governed largely by the promoter used to express the gene. A promoter is a DNA sequence that directs the cellular machinery of a plant to produce (transcribe) RNA (transcript) from a contiguous transcribable region downstream (3') of the promoter. The promoter influences the rate at which the transcript of the gene is made. Assuming the transcript includes a coding region with appropriate translational signals, the promoter also influences the rate at which the resultant protein product of the gene is produced. Promoter activity also can depend on the presence of several other *cis*-acting regulatory elements which, in conjunction with cellular factors, determine strength, specificity, and transcription initiation site (for a review, see Zawel and Reinberg, 1992, *Curr. Opin. Cell Biol.* 4:488).

It has been shown that certain promoters are able to direct RNA synthesis at a higher rate relative to other promoters. These are called "strong promoters". Certain other promoters have been shown to direct RNA production at higher levels only in particular types of cells or tissues and are often referred to as "tissue-specific promoters". Promoters that are capable of directing RNA production in many or all tissues of a plant are called "constitutive promoters". Thus, expression of a chimeric gene (or genes) introduced into a plant may potentially be controlled by identifying and using a promoter with the desired characteristics.

A myriad of promoters has been described for gene expression in dicotyledonous plants. However, there is currently a dearth of promoters that can be used for effective expression of foreign or endogenous coding sequences in monocotyledonous plants.

SUMMARY OF THE INVENTION

Thus, in one aspect, the invention provides an isolated nucleic acid comprising a nucleotide sequence which corresponds to a promoter region of a transcribable DNA sequence that is hybridizable to a probe or primer derivable from a polynucleotide sequence selected from the group consisting of: -

- (a) the polynucleotide sequence set forth in FIG. 14 [SEQ ID NO: 1];

- (b) the polynucleotide sequence set forth in FIG. 15 under designator c51 [SEQ ID NO: 2];
- (c) the polynucleotide sequence set forth in FIG. 15 under designator c511 [SEQ ID NO: 3];
- 5 (d) the polynucleotide sequence set forth in FIG. 15 under designator c512 [SEQ ID NO: 4];
- (e) the polynucleotide sequence set forth in FIG. 16 [SEQ ID NO: 5];
- (f) the polynucleotide sequence set forth in FIG. 17 [SEQ ID NO: 6];
- (g) the polynucleotide sequence set forth in FIG. 23 [SEQ ID NO: 7];
- 10 (h) the polynucleotide sequence set forth in FIG. 24 [SEQ ID NO: 8];
- (i) the polynucleotide sequence set forth in FIG. 26 [SEQ ID NO: 9];
- (j) the polynucleotide sequence set forth in FIG. 29 [SEQ ID NO: 10]; and
- (k) the polynucleotide sequence set forth in FIG. 31 [SEQ ID NO: 11].

Advantageously, the isolated promoter region of the transcribable
15 DNA sequence is of sufficient length such that it is capable of initiating and regulating transcription of a DNA sequence to which it is coupled. The promoter region may be between 100 bp and 4 kb in length and preferably greater than 1 kb in length. An analogous promoter region can be obtained from any plant species that has a DNA sequence that is transcribed in one or more cells or tissues of the plant
20 provided that the DNA sequence is capable of hybridising to a nucleic acid probe derivable from a polynucleotide sequence as set forth in any one of SEQ ID NO: 1, 2, 3, 4, 5, 6, 7, 8, 9, 10 and 11 under at least low stringency conditions, preferably under at least medium stringency conditions, and more preferably under high stringency conditions.

25 The polynucleotide sequences set forth in SEQ ID NO: 1, 2, 3, 4, 5, 6, 7, 8, 9, 10 and 11 are transcribable sequences obtained from sugarcane (*Saccharum* sp.). The polynucleotide sequences identified by SEQ ID NO: 1, 2, 3, 4, 9, 10 and 11 are highly transcribed in stem tissue, particularly mature stem tissue of sugarcane. Homologous sequences corresponding to SEQ ID NO: 2, 3 and 4 are expressed in

stem tissue of other monocotyledonous plants including maize and sorghum. Homologous sequences corresponding to SEQ ID NO: 1, 9, 10 and 11 are also suspected to exist in other monocotyledonous plants.

5 The invention thus also provides a polynucleotide sequence or variant thereof that is highly transcribed in stem tissue of monocotyledonous plants wherein said sequence is selected from the group consisting of any one of the polynucleotides set forth in SEQ ID NO: 1, 2, 3, 4, 9, 10 and 11.

10 The polynucleotide sequences identified by SEQ ID NO: 7 and 8 are highly transcribed in meristem tissue of sugarcane. Thus, the invention also provides a polynucleotide sequence or variant thereof that is highly transcribed in meristem tissue of monocotyledonous plants wherein said sequence is selected from the group consisting of SEQ ID NO: 7 or SEQ ID NO: 8.

15 The polynucleotides identified by SEQ ID NO: 5 and 6 are expressed constitutively in leaves, stems and roots of sugarcane. Homologous sequences corresponding to these polynucleotides are expressed in many or all tissues of other monocotyledonous plants including maize, rice and sorghum.

20 Accordingly, the invention also features a polynucleotide sequence or variant thereof that is highly transcribed constitutively in tissue of monocotyledonous plants wherein said sequence is selected from the group consisting of: -

(a) the polynucleotide sequence set forth in SEQ ID NO: 5; and

(b) the polynucleotide sequence set forth in SEQ ID NO: 6.

25 The foregoing polynucleotide sequences represent transcribable sequences, which can be used to construct a probe or primer(s) to isolate homologous transcribable sequences in other plant species, preferably, monocotyledonous species, so that a corresponding promoter region from said other plant species having the same tissue-specific or constitutive qualities can be isolated and used.

30 Preferably, the isolated nucleic acid comprises a nucleotide sequence selected from the group consisting of:

- (a) the polynucleotide sequence set forth in FIG. 18 [SEQ ID NO: 12];
- (b) the polynucleotide sequence set forth in FIG. 19 [SEQ ID NO: 13];
- (c) the polynucleotide sequence set forth in FIG. 20 [SEQ ID NO: 14];
- (d) the polynucleotide sequence set forth in FIG. 21 [SEQ ID NO: 15];
- 5 (e) a biologically-active portion of any one of SEQ ID NO: 12, 13, 14 or 15; and
- (f) a variant of any one of the polynucleotide sequences according to (a) to (e).

In one embodiment, the variant has at least 60%, preferably at least 80%, more preferably at least 90%, and most preferably at least 95% sequence identity to any one of the polynucleotides set forth in SEQ ID NO: 12, 13, 14 or 15.

10 In another embodiment, the variant is capable of hybridising to any one of the polynucleotides identified by SEQ ID NO: 12, 13, 14 or 15 under at least low stringency conditions, preferably under at least medium stringency conditions, and more preferably under high stringency conditions.

15 In another aspect, the invention provides a chimeric gene comprising an isolated nucleic acid of the invention, variant or biologically-active fragment and a heterologous nucleic acid.

In yet another aspect, the invention provides an expression vector comprising an isolated nucleic acid of the invention, variant or biologically-active fragment.

20 Preferably, the expression vector comprises an isolated nucleic acid of the invention, variant or biologically-active fragment and a heterologous nucleic acid operably linked thereto.

In one embodiment, the heterologous nucleic acid encodes a polypeptide such as a structural or regulatory protein.

25 In another embodiment, the heterologous nucleic acid encodes a molecule which is capable of modulating expression of a target gene.

In a preferred embodiment, the molecule is an antisense RNA or a ribozyme or other transcribed region aimed at downregulation of expression of the corresponding target gene. For example, the said other transcribed region may

comprise a sense transcript aimed at sense suppression (co-suppression) of the target gene.

Depending on the polynucleotide selected, in one embodiment, the expression construct may be further characterized in that said isolated nucleic acid is capable of directing transcription of the heterologous nucleic acid preferentially in stem tissue of a plant. In another embodiment, expression construct may be further characterized in that that said isolated nucleic acid is capable of directing transcription of the heterologous nucleic acid preferentially in meristem tissue of a plant. In yet another embodiment, the expression construct may be further characterized in that said isolated nucleic acid is capable of directing transcription of the heterologous nucleic acid in many or all tissues of a plant.

In a further aspect, the invention provides a method of transforming a plant, including the step of introducing into a plant cell or tissue an isolated nucleic acid, chimeric gene or expression vector of the invention.

According to a still further another aspect of the invention, there is provided a transformed plant cell or tissue comprising an isolated nucleic acid, chimeric gene or expression vector of the invention.

In a still yet further aspect, the invention provides a transformed plant comprising an isolated nucleic acid, chimeric gene or expression vector of the invention.

Plants encompass any taxonomic grouping thereof, including angiosperms, gymnosperms, monocotyledons and dicotyledons. Preferred plants are monocotyledons such as cereals, sugarcane, bananas and pineapples, but without limitation thereto.

More preferably, the plant is sugarcane.

Preferably, the transformed plant has an altered phenotype compared to a corresponding non-transformed plant.

Preferably, the altered phenotype results from expression of the heterologous nucleic acid.

The invention also provides cells, tissues, leaves, fruit, flowers, seeds and other reproductive material, material used for vegetative propagation, progeny plants including F1 hybrids, male-sterile plants and all other plants and plant products derivable from transgenic plants of the invention.

5

BRIEF DESCRIPTION OF THE DRAWINGS

In order that the invention may be readily understood and put into practical effect, preferred embodiments will now be described by way of example with reference to the accompanying drawings in which:

FIG. 1: Northern analysis of total RNA extracted from leaf (L), stem
10 (S) and root (R) of field-grown sugarcane plants. The cultivars were Pindar, NCo310, Q110 and Q145. The probe is the cDNA c67 (SEQ ID NO: 1), revealing preferential expression of gene 67 in the stems of all tested sugarcane cultivars. The relative expression levels for the leaf / stem / root tissues are 0.7 / 100 / 5 (Pindar), 0.3 / 100 / 1.7 (NCo310), 0.3 / 100 / 2.8 (Q110) and 0.7 / 100 / 2 (Q145).

15 FIG. 2: Northern analysis of total RNA extracted from different internodes of field-grown sugarcane plants (cultivar Pindar). Each plant had a total of 17 internodes, IN₁ corresponds to internode 1 (top of the stem) with other internodes numbered down the stem. The probe is the cDNA c67 (SEQ ID NO: 1), revealing preferential expression of gene 67 in mature internodes. The relative
20 expression levels are 0 / 0.7 / 9.7 / 47 / 100 / 45 (IN₁ / IN₄ / IN₆ / IN₈ / IN₁₁ / IN₁₅).

FIG. 3: Northern analysis of total RNA extracted from leaf (L), stem
(S) and root (R) of field-grown sugarcane plants. The cultivars were Pindar, NCo310, Q110 and Q145. The probe is a *NheI* DNA fragment from the cDNA c51
(SEQ ID NO: 2, 3 and 4), revealing stem-preferential expression of gene 51 in all
25 tested sugarcane cultivars. The relative expression levels for the leaf / stem / root tissues are 19.5 / 100 / 18 (Pindar), 17.5 / 100 / 23.7 (NCo310), 16.8 / 100 / 20 (Q110) and 12.5 / 100 / 15.8 (Q145).

FIG. 4: Northern analysis of total RNA extracted from different parts
of cultivar Pindar sugarcane plants with 17 internodes. IN₆ and IN₁₁ correspond
30 respectively to internode 6 and 11. Nc is the core of a nodal part of the stem and N is

an entire nodal part of the stem. M corresponds to the meristematic region of the stem. R is the root tissue. YL, ML and OL represent different stages of leaf development, Y being the young and recently emerged leaves, M being the more mature leaves and O being leaves at an older stage. The probe is the cDNA c51
 5 (SEQ ID NO: 2, 3 and 4), revealing expression of gene 51 throughout the sugarcane stem.

FIG. 5: Northern analysis of total RNA extracted from different parts of field-grown maize plants and glasshouse-grown sorghum plants. The different tissues analysed were male flower parts (F), top of the stem (TS), medium and lower
 10 part of the stem (MS and LS), root system (R), leaves at a young, mature and older stage of development (YL, ML and OL) and panicle (P). The probe is the cDNA c51, revealing expression of a homologue of sugarcane stem-specific gene 51 (SEQ ID NO: 2, 3 and 4) in maize and sorghum.

FIG. 6: Northern analyses of total RNA extracted from different parts
 15 of field-grown sugarcane (cultivar Pindar) and maize (sweet corn) stems. For the sugarcane RNA, each plant had a total of 17 internodes, IN₁ corresponds to internode 1 (top of the stem), with other internodes numbered down the stem. For the maize RNA, each plant had a total of 13 internodes, IN₁ corresponds to internode 1 (top of the stem), with other internodes numbered down the stem. The probe is the cDNA
 20 c51 (SEQ ID NO: 2, 3 and 4), revealing expression of gene 51 in internodes at all stages of maturity in both sugarcane and maize. The relative expression levels are 44 / 47 / 51 / 64 / 100 / 61 (IN₁ / IN₄ / IN₆ / IN₈ / IN₁₁ / IN₁₅) in sugarcane stems and 40 / 44 / 75 / 87 / 100 / 72 (IN₁ / IN₃ / IN₅ / IN₇ / IN₉ / IN₁₁) in maize stems.

FIG. 7: Northern analysis of total RNA extracted from various parts
 25 of field-grown sugarcane plants (cultivar Pindar). Each plant had a total of 17 internodes. The tissues analysed were a nodal part of the stem (N), internodes 1-2 (IN₁₋₂, top of the stem), 6 and 11 (IN₆ and IN₁₁), the root system (R) and leaves at a young, mature and older stage of development (YL, ML and OL). The probe is the cDNA c32A (SEQ ID NO: 5 and 6), revealing expression of gene 32A in all tested
 30 sugarcane tissues. The relative expression levels are 42 / 100 / 92 / 94 / 59 / 43 / 70 / 60 (N / IN₁₋₂ / IN₆ / IN₁₁ / R / YL / ML / OL).

FIG. 8: Northern analysis of total RNA extracted from leaf (L), stem (S) and root (R) of field-grown sugarcane plants. The cultivars were Pindar, NCo310, Q110 and Q145. The probe is the cDNA c322 (SEQ ID NO: 5 and 6), revealing constitutive expression of gene 32A in all tested sugarcane cultivars. The relative expression levels for the leaf / stem / root tissues are 58 / 100 / 40 (Pindar), 48 / 100 / 43 (NCo310), 88 / 100 / 58 (Q110) and 70 / 100 / 51 (Q145).

FIG. 9: Northern analyses of total RNA extracted from different parts of field-grown sugarcane (cultivar Pindar) and maize (sweet corn) stems. For the sugarcane RNA, each plant had a total of 17 internodes, IN₁ corresponds to internode 1 (top of the stem), with other internodes numbered down the stem. For the maize RNA, each plant had a total of 13 internodes, IN₁ corresponds to internode 1 (top of the stem), with other internodes numbered down the stem. The probe is the cDNA c322 (SEQ ID NO: 5 and 6), revealing expression of gene 32A and its maize homologue in internodes at all stages of maturity in both sugarcane and maize. The relative expression levels are 81 / 71 / 81 / 98 / 100 / 74.5 (IN₁ / IN₄ / IN₆ / IN₈ / IN₁₁ / IN₁₅) in sugarcane stems and 74.5 / 94 / 97.5 / 100 / 82 / 90 (IN₁ / IN₃ / IN₅ / IN₇ / IN₉ / IN₁₁) in maize stems.

FIG. 10: Northern analysis of total RNA extracted from glasshouse-grown sorghum and rice plants. For the sorghum RNA, the different tissues analysed were the panicle (P), the top, middle part and lower parts of the stem (TS, MS and LS), the root system (R) and leaves at a young, mature and older stage of development (YL, ML and OL). For the rice RNA, the different tissues analysed were the young and older stems (S₂ and S₁), the root system (R) and the young, mature and older leaves (L₃, L₂ and L₁). The probe is the cDNA c322 (SEQ ID NO: 5 and 6), revealing a constitutive expression of homologues of gene 32A in all the tissues tested in sorghum and rice.

FIG. 11: Southern analysis of genomic DNA (10 µg/lane) from cultivar Q117 digested by *Eco*RI (E), *Kpn*I (K), *Sac*I (S), *Xba*I (X) and *Not*I (N), indicating genomic copies of gene 67 in sugarcane. The probe is the cDNA c67 (SEQ ID NO: 1). Lane λ_{H3} indicates the position of molecular weight markers on the same electrophoresis gel.

FIG. 12: Southern analysis of genomic DNA (10 µg/lane) from cultivar Q117 digested by *EcoRI* (E), *KpnI* (K), *SacI* (S), *XbaI* (X) and *NotI* (N), indicating few genomic copies of gene 51 in sugarcane. The probe is the 3' region of the cDNA c51 (nucleotide 621 to nucleotide 874 of SEQ ID NO: 2, 3 and 4). Lane 5 λ_{H3} indicates the position of molecular weight markers on the same electrophoresis gel.

FIG. 13: Southern analysis of genomic DNA (10 µg/lane) from cultivar Q117 digested by *EcoRI* (E), *KpnI* (K), *SacI* (S), *XbaI* (X) and *NotI* (N), indicating multiple copies of gene 32A in sugarcane. The probe is the cDNA c322 (SEQ ID NO: 5 and 6). Lane 10 λ_{H3} indicates the position of molecular weight markers on the same electrophoresis gel.

FIG. 14: Nucleotide sequence of the cDNA c67 (SEQ ID NO: 1) and deduced amino acid sequence of the longest open reading frame (SEQ ID NO: 16). The ATG start codon is at position 33 to 35, the stop codon (TGA) is at position 594 to 596. No homologous sequences (with more than 40% identity on the whole sequence) were found in the databases (NR nucleic, ANGIS) at either the nucleotide or the amino acid level. Two putative polyadenylation signals (AATAAA) are present between nucleotide 853 and nucleotide 862.

FIG. 15: Nucleotide sequence alignment of the cDNA clones c51, c511 and c512 (SEQ ID NO: 2, 3 and 4) and deduced amino acid sequence (aa) of the longest and conserved open reading frames (SEQ ID NO: 17). The ATG start codon has been assigned to position 129 to 131, the stop codon (TGA) is at position 576 to 578. The cDNA clone c511 has a stop codon TAG at position 435 to 437. The alanine (A) residue deduced from nucleotide position 402-404 in the cDNAs c51 and c511 is changed to a valine residue (V) in the deduced amino acid sequence from the cDNA c512. The valine (V) residue deduced from nucleotide position 501-503 in the cDNAs c511 and c512 is changed to an alanine (A) residue in the deduced amino acid sequence of the cDNA c51. The nucleotide and deduced amino-acid sequences of all the cDNAs have no homologues in the nucleotide and protein databases (NR nucleic, ANGIS).

FIG. 16: Nucleotide sequence of cDNA clone c32A (SEQ ID NO: 5) and deduced amino acid sequence (SEQ ID NO: 22). The stop codon (TAG) is at position 44 to 46. This partial protein sequence (SEQ ID NO: 22) is homologous to S27 ribosomal proteins from rice, barley, *Arabidopsis thaliana*, *Chlamydomonas reinhardtii* and other organisms.

FIG. 17: Nucleotide sequence and deduced amino acid sequence of the cDNA clone c322 (SEQ ID NO: 6 and 23, respectively). The ATG start codon is at position 92 to 94, the stop codon (TAG) is at position 350 to 352. The protein sequence (SEQ ID NO: 23) is homologous to S27 ribosomal proteins from rice, barley, *Arabidopsis thaliana*, *Chlamydomonas reinhardtii* and other organisms.

FIG. 18: Nucleotide sequence of the 67pro promoter sequence (SEQ ID NO: 12). The start of the coding sequence is at position 1045. A putative "tata box" is present at position 964-969. The predicted start of transcription is at position 994 (A).

FIG. 19: Nucleotide sequence of the 32a2pro promoter sequence (SEQ ID NO: 13). The start of the coding sequence is at position 1943.

FIG. 20: Nucleotide sequence of the 32a6pro promoter sequence (SEQ ID NO: 14). The start of the coding sequence is at position 1266. A putative "tata box" is present at position 1138-1145. The predicted start of transcription is at position 1170 (C).

FIG. 21: Nucleotide sequence of the 51pro promoter sequence (SEQ ID NO: 15). The start of the coding sequence is at position 2946.

FIG. 22: GUS histochemical assay of cross-sections of three transgenic sugarcane plant stems transformed with plasmid p67G. The strong blue staining (shown as dark grey to black on the figure) indicates the presence of the GUS protein and confirms the functionality of the promoter fused to the GUS gene.

FIG. 23: Nucleotide sequence of the cDNA c19 (SEQ ID NO: 7). Neither the nucleotide sequence nor the deduced amino acid sequences (in the six frames) have homologues in the databases (NR nucleic and NR Proteins, ANGIS).

FIG. 24: Nucleotide sequence of the cDNA c3 (SEQ ID NO: 8). The sequence from nucleotide 144 to nucleotide 395 (deduced amino acid sequence SEQ ID NO: 21) is homologous to part of the cellulase (EC 3.2.1.4) coding sequence from a variety of sources (e.g. *Arabidopsis thaliana*, tomato, tobacco, pepper, termites and bacteria).

FIG. 25: Northern analysis of total RNA extracted from different parts of field-grown sugarcane plants. The probe is the cDNA c19 (SEQ ID NO: 7), revealing preferential expression in the meristematic region of the stem (M). No expression can be detected in other tissues such as the core region of a stem internode (INc), a whole stem internode (IN), the core region of a stem node (Nc), a whole stem node (N), the root system (R), young leaves (YL), mature leaves (ML) and older leaves (OL). An identical result is obtained (except for the size of the hybridising transcript) when using the cDNA c3 (SEQ ID NO: 8) as the probe.

FIG. 26: Nucleotide sequence and deduced amino acid sequence of the cDNA c18 (SEQ ID NO: 9 and 18, respectively). The ATG start codon is present at position 1-3 and the stop codon (TGA) is at position 244-246. The amino acid sequence is homologous to type 2 plant metallothionein-like proteins previously described in different plants such as *Arabidopsis thaliana*, tobacco, barley, fava bean and others.

FIG. 27: Northern analysis of total RNA extracted from leaf (L), stem (S) and root (R) of field-grown sugarcane plants. The cultivars were Pindar, NCo310, Q110 and Q145. The probe is the cDNA c18 (SEQ ID NO: 9), revealing preferential expression of gene 18 in the stems of all tested sugarcane cultivars. The relative expression levels for the leaf / stem / root tissues are 6 / 100 / 8.5 (Pindar), 9 / 100 / 10 (NCo310), 11.5 / 100 / 11.5 (Q110) and 6 / 100 / 5 (Q145).

FIG. 28: Northern analysis of total RNA extracted from different internodes of field-grown sugarcane plants (cultivar Pindar). Each plant had a total of 17 internodes, IN₁ corresponds to internode 1 (top of the stem) with other internodes numbered down the stem. The probe is the cDNA c18 (SEQ ID NO: 9), revealing preferential expression in the young parts of the stem, with a gradual decrease in the expression levels towards the base of the stem (older parts). The

relative expression levels are 100 / 98 / 86 / 69 / 40 / 25 (IN₁ / IN₄ / IN₆ / IN₈ / IN₁₁ / IN₁₅).

FIG. 29: Nucleotide sequence of the cDNA c53A (SEQ ID NO: 10) and deduced amino acid sequence (SEQ ID NO: 19). The ATG start codon is at position 124-126, the stop codon TGA is at position 1069-1071. The protein sequence is homologous to homeobox proteins described in plants (e.g. *Arabidopsis thaliana* ATK1 protein and KNA2 knotted-like homeobox protein, soybean HMB1 homeobox protein, maize RS1 gene product, rice OSH1 homeobox 1 protein and others).

FIG. 30: Northern analysis of total RNA extracted from leaf (L), stem (S) and root (R) of field-grown sugarcane plants. The cultivars were Pindar, Nco310, Q110 and Q145. The probe is the cDNA c53A (SEQ ID NO: 10), revealing preferential expression of gene 53A in the stems of all tested sugarcane cultivars. The relative expression levels for the leaf / stem / root tissues are 4 / 100 / 14 (Pindar), 5 / 100 / 12 (NCo310), 0 / 100 / 11 (Q110) and 0 / 100 / 8 (Q145).

FIG. 31: Nucleotide sequence (SEQ ID NO: 11) and deduced amino acid sequence (SEQ ID NO: 20) of the cDNA c57. The stop codon (TGA) is at position 567-569. The amino acid sequence and the nucleotide coding sequence are homologous to several auxin-induced and auxin-responsive genes / proteins from different plants such as *Arabidopsis thaliana*, soybean and tobacco.

FIG. 32: Northern analysis of total RNA extracted from leaf (L), stem (S) and root (R) of field-grown sugarcane plants. The cultivars were Pindar, Nco310, Q110 and Q145. The probe is the cDNA c57 (SEQ ID NO: 11), revealing preferential expression of gene 57 in the stems of all tested sugarcane cultivars. The relative expression levels for the leaf / stem / root tissues are 0 / 100 / 0 (Pindar), 0 / 100 / 0 (Nco310), 15 / 100 / 7 (Q110) and 15 / 100 / 12 (Q145).

FIG. 33: Northern analysis of total RNA extracted from different internodes of field-grown sugarcane plants (cultivar Pindar). Each plant had a total of 17 internodes, IN₁ corresponds to internode 1 (top of the stem) with other internodes numbered down the stem. The probe is the cDNA c57 (SEQ ID NO: 11), revealing preferential expression in the young parts of the stem, with a gradual

decrease in the expression levels towards the base of the stem (older parts). The relative expression levels are 100 / 86 / 82 / 65 / 45 / 20 (IN₁ / IN₄ / IN₆ / IN₈ / IN₁₁ / IN₁₅).

BRIEF DESCRIPTION OF THE SEQUENCES

- 5 SEQ ID NO: 1 Nucleotide sequence of cDNA clone c67,
which is preferentially expressed in the stem of field-grown sugarcane plants.
- SEQ ID NO: 2 Nucleotide sequence of cDNA clone c51,
which is preferentially expressed in the stem of field-grown sugarcane plants.
- SEQ ID NO: 3 Nucleotide sequence of cDNA clone c511,
10 which is preferentially expressed in the stem of field-grown sugarcane plants.
- SEQ ID NO: 4 Nucleotide sequence of cDNA clone c512,
which is preferentially expressed in the stem of field-grown sugarcane plants.
- SEQ ID NO: 5 Nucleotide sequence of cDNA clone c32A,
which is transcribed constitutively in leaves, stems and roots of field-grown
15 sugarcane plants.
- SEQ ID NO: 6 Nucleotide sequence of cDNA clone c322,
which is transcribed constitutively in leaves, stems and roots of field-grown
sugarcane plants.
- SEQ ID NO: 7 Nucleotide sequence of cDNA clone c19,
20 which is preferentially expressed in meristematic tissue of field-grown sugarcane
plants.
- SEQ ID NO: 8 Nucleotide sequence of the cDNA c3, which is
preferentially expressed in meristematic tissue of field-grown sugarcane plants.
- SEQ ID NO: 9 Nucleotide sequence of cDNA clone c18,
25 which is preferentially expressed in the stem of field-grown sugarcane plants.
- SEQ ID NO: 10 Nucleotide sequence of cDNA clone c53A,
which is preferentially expressed in the stem of field-grown sugarcane plants.
- SEQ ID NO: 11 Nucleotide sequence of cDNA clone c57,
which is preferentially expressed in the stem of field-grown sugarcane plants.

SEQ ID NO: 12 Nucleotide sequence of the stem-specific 67 promoter. *E. coli* DH10B strain transformed with plasmid pG4-67pro comprising the 67 promoter was deposited with AGAL on August 30, 1999 under accession number NM99/05995.

5 SEQ ID NO: 13 Nucleotide sequence of the stem-specific 32A2 promoter. *E. coli* TOP10 strain transformed with pZ-32A2P2 comprising the 32A2 promoter was deposited with AGAL on August 30, 1999 under accession number NM99/05994.

10 SEQ ID NO: 14 Nucleotide sequence of the constitutive 32A6 promoter. *E. coli* TOP10 strain transformed with pZ-32A6P2 comprising the 32A6 promoter was deposited with AGAL on August 30, 1999 under accession number NM99/05993.

15 SEQ ID NO: 15 Nucleotide sequence of the constitutive 51 promoter. *E. coli* TOP10 strain transformed with pZ-H51-6P1P2 comprising the 51 promoter was deposited with AGAL on August 30, 1999 under accession number NM99/05992.

SEQ ID NO: 16 Deduced amino acid sequence of the longest open reading frame of SEQ ID NO: 1.

20 SEQ ID NO: 17 Deduced amino acid sequence of the longest and conserved open reading frame common to each of SEQ ID NO: 2, 3 and 4.

SEQ ID NO: 18 Deduced amino acid sequence of the longest open reading frame of SEQ ID NO: 5.

SEQ ID NO: 19 Deduced amino acid sequence of the longest open reading frame of SEQ ID NO: 10.

25 SEQ ID NO: 20 Deduced amino acid sequence of the longest open reading frame of SEQ ID NO: 11.

SEQ ID NO: 21 Deduced amino acid sequence of the longest open reading frame of SEQ ID NO: 8.

SEQ ID NO: 22 Deduced amino acid sequence of the longest
open reading frame of SEQ ID NO: 5.

SEQ ID NO: 23 Deduced amino acid sequence of the longest
open reading frame of SEQ ID NO: 6.

5

DETAILED DESCRIPTION

1. Definitions

Unless defined otherwise, all technical and scientific terms used herein have the same meaning as commonly understood by those of ordinary skill in the art to which the invention belongs. Although any methods and materials similar or equivalent to those described herein can be used in the practice or testing of the present invention, preferred methods and materials are described. For the purposes of the present invention, the following terms are defined below.

10

"Amplification product" refers to a nucleic acid product generated by nucleic acid amplification techniques.

15

By *"biologically active fragment"* is meant a segment, portion or fragment of a the biological active molecule which has at least about 0.1%, preferably at least about 10%, more preferably at least about 25% and even more preferably at least 50% of the activity of the molecule.

20

"Chimeric gene" is defined herein as a nucleic acid, preferably a DNA molecule, either single- or double-stranded, which includes an isolated nucleic acid of the invention, variant or biologically-active fragment, operably linked to a heterologous nucleic acid.

25

By *"corresponds to"* or *"corresponding to"* is meant similar, related or analogous to in structure and/or function. For example, an isolated nucleic acid of the invention suitably comprises a nucleotide sequence structurally and/or functionally related to a corresponding promoter sequence, such that the isolated nucleic acid of the invention is capable of functioning as a promoter.

The term "*regeneration*" as used herein means growing a whole, differentiated plant from a plant cell, a group of plant cells, a plant part (including seeds), or a plant piece (e.g., from a protoplast, callus, or tissue part).

5 By "*heterologous nucleic acid*" is meant a nucleic acid distinct from an isolated promoter of the invention. Operationally, the heterologous nucleic acid is operably linked to an isolated promoter nucleic acid of the invention to achieve expression of the heterologous nucleic acid. The term heterologous nucleic acid encompasses transcribable DNA as will be defined hereinafter.

10 "*Homology*" refers to the percentage number of nucleotides of a polynucleotide sequence that are identical to a reference polynucleotide sequence. Homology may be determined using sequence comparison programs such as BESTFIT (Deveraux *et al.* 1984, *Nucleic Acids Research* 12, 387-395) which is incorporated herein by reference. In this way sequences of a similar or substantially different length to those cited herein might be compared by insertion of gaps into the
15 alignment, such gaps being determined, for example, by the comparison algorithm used by BESTFIT.

"*Hybridization*" is used herein to denote the pairing of complementary nucleotide sequences to produce a DNA-DNA hybrid or a DNA-RNA hybrid. Complementary base sequences are those sequences that are related by the base-
20 pairing rules. In DNA, A pairs with T and C pairs with G. In RNA U pairs with A and C pairs with G. In this regard, the terms "match" and "mismatch" as used herein refer to the hybridisation potential of paired nucleotides in complementary nucleic acid strands. Matched nucleotides hybridise efficiently, such as the classical A-T and G-C base pair mentioned above. Mismatches are other combinations of nucleotides
25 that do not hybridise efficiently.

By "*isolated*" is meant material that is substantially or essentially free from components that normally accompany it in its native state. For example, an "isolated nucleic acid", as used herein, refers to a nucleic acid, which has been purified from the sequences which flank it in a naturally occurring state, e.g., a DNA
30 fragment which has been removed from the sequences which are normally adjacent to the fragment.

By "*marker gene*" is meant a gene that imparts a distinct phenotype to cells expressing the marker gene and thus allows such transformed cells to be distinguished from cells that do not have the marker. A selectable marker gene confers a trait for which one can 'select' based on resistance to a selective agent (e.g., a herbicide, antibiotic, radiation, heat, or other treatment damaging to untransformed cells). A screenable marker gene (or reporter gene) confers a trait that one can identify through observation or testing, i.e., by 'screening' (e.g. green fluorescent protein or enzymes such as β -glucuronidase, neomycin phosphotransferase II, and luciferase not present in untransformed cells).

10 The term "*nucleic acid*" as used herein designates DNA and/or RNA including mRNA, cRNA, cDNA and genomic DNA in single-stranded and double-stranded form, and encompasses oligonucleotides and polynucleotides as herein defined.

15 By "*obtained from*" is meant that a sample such as, for example, a nucleic acid extract is isolated from, or derived from, a particular source of the host. For example, the nucleic acid extract may be obtained from tissue isolated directly from the host.

20 The term "*oligonucleotide*" as used herein refers to a polymer composed of a multiplicity of nucleotide units (deoxyribonucleotides or ribonucleotides, or related structural variants or synthetic analogues thereof) linked via phosphodiester bonds (or related structural variants or synthetic analogues thereof). Thus, while the term "*oligonucleotide*" typically refers to a nucleotide polymer in which the nucleotides and linkages between them are naturally occurring, it will be understood that the term also includes within its scope various analogues including, but not restricted to, peptide nucleic acids (PNAs), phosphoramidates, phosphorothioates, methyl phosphonates, 2-O-methyl ribonucleic acids, and the like. The exact size of the molecule may vary depending on the particular application. An oligonucleotide typically comprises from about 10 to 30 nucleotides, but the term can refer to molecules of any length, although the term "*polynucleotide*" usually refers to nucleic acids of more than 100 nucleotides in length.

30 By "*operably linked*" is meant functionally linked. For example, an

isolated nucleic acid of the invention is operably linked to a heterologous nucleic acid regulated by being linked thereto so as to be capable of initiating, controlling, regulating or otherwise directing transcription of the heterologous nucleic acid. Usually, the isolated nucleic acid is located upstream or 5' of the heterologous nucleic acid.

"Polypeptide", "peptide" and "protein" are used interchangeably herein to refer to a polymer of amino acid residues and to variants and synthetic analogues of the same. Thus, these terms apply to amino acid polymers in which one or more amino acid residues is a synthetic non-naturally occurring amino acid, such as a chemical analogue of a corresponding naturally occurring amino acid, as well as to naturally-occurring amino acid polymers.

By "primer" is meant an oligonucleotide or polynucleotide which is capable of hybridizing to a complementary or partly complementary nucleic acid strand (template) and initiating synthesis of a primer extension product in the presence of a suitable polymerising agent. The primer is preferably single-stranded for maximum efficiency in amplification but may alternatively be double-stranded. A primer must be sufficiently long to prime the synthesis of extension products in the presence of the polymerisation agent. The length of the primer depends on many factors, including application, temperature to be employed, template reaction conditions, other reagents, and source of primers. For example, depending on the complexity of the target sequence, the oligonucleotide primer typically contains 15 to 35 or more nucleotides, although it may contain fewer nucleotides. Primers can be large polynucleotides, such as from about 200 nucleotides to several kilobases or more. Primers may be selected to be "substantially complementary" to the sequence on the template to which it is designed to hybridise and serve as a site for the initiation of synthesis. By "substantially complementary", it is meant that the primer is sufficiently complementary to hybridise with the template. Preferably, the primer contains no mismatches with the template to which it is designed to hybridise but this is not essential. For example, non-complementary nucleotides may be attached to the 5'-end of the primer, with the remainder of the primer sequence being complementary to the template. Alternatively, non-complementary nucleotides or a stretch of non-complementary nucleotides can be interspersed into a primer,

provided that the primer sequence has sufficient complementarity with the sequence of the template to hybridise therewith and thereby form a template for synthesis of the extension product of the primer.

5 "*Probe*" refers to a molecule that binds to a specific sequence or sub-sequence or other moiety of another molecule. Unless otherwise indicated, the term "probe" typically refers to a polynucleotide probe that binds to another nucleic acid, often called the "target nucleic acid", through complementary base pairing. Probes may bind target nucleic acids lacking complete sequence complementarity with the probe, depending on the stringency of the hybridisation conditions. Probes can be
10 directly or indirectly labelled.

The term "*promoter*" refers to a nucleic acid which directs expression of another nucleic acid to which it is operably linked, by initiating, regulating or otherwise controlling transcription of said another nucleic acid.

15 "*Constitutive promoter*" refers to a promoter that directs transcription in many or all tissues of a plant.

By "*stem-specific promoter*" is meant a promoter that preferentially directs transcription in stem tissue of a plant.

By "*meristem-specific promoter*" is meant a promoter that preferentially directs transcription in meristem tissue of a plant.

20 The term "*recombinant*" as used herein refers to a nucleic acid or polypeptide resulting from *in vitro* manipulation into a form not normally found in nature. As used in the art, "*recombinant*" usually refers to the products of recombinant DNA technology.

25 Terms used to describe sequence relationships between two or more polynucleotides or polypeptides include "reference sequence", "comparison window", "sequence identity", "percentage of sequence identity" and "substantial identity". A "*reference sequence*" is at least 6 but frequently 15 to 18 and often at least 25 monomer units, inclusive of nucleotides and amino acid residues, in length. Because two polynucleotides may each comprise (1) a sequence (*i.e.*, only a portion
30 of the complete polynucleotide sequence) that is similar between the two

polynucleotides, and (2) a sequence that is divergent between the two polynucleotides, sequence comparisons between two (or more) polynucleotides are typically performed by comparing sequences of the two polynucleotides over a "comparison window" to identify and compare local regions of sequence similarity.

- 5 A "comparison window" refers to a conceptual segment of typically 6 to 12 contiguous residues that is compared to a reference sequence. The comparison window may comprise additions or deletions (*i.e.*, gaps) of about 20% or less as compared to the reference sequence (which does not comprise additions or deletions) for optimal alignment of the two sequences. Optimal alignment of sequences for
10 aligning a comparison window may be conducted by computerised implementations of algorithms (GAP, BESTFIT, FASTA, and TFASTA in the Wisconsin Genetics Software Package Release 7.0, Genetics Computer Group, 575 Science Drive Madison, WI, USA, incorporated herein by reference) or by inspection and the best alignment (*i.e.*, resulting in the highest percentage homology over the comparison
15 window) generated by any of the various methods selected. Reference also may be made to the BLAST family of programs as for example disclosed by Altschul *et al.*, 1997, *Nucl. Acids Res.* 25:3389, which is incorporated herein by reference. A detailed discussion of sequence analysis can be found in Unit 19.3 of Ausubel *et al.*, "Current Protocols in Molecular Biology", John Wiley & Sons Inc, 1994-1998,
20 Chapter 15.

- The term "sequence identity" as used herein refers to the extent that sequences are identical on a nucleotide-by-nucleotide basis over a window of comparison. Thus, a "percentage of sequence identity" is calculated by comparing two optimally aligned sequences over the window of comparison, determining the
25 number of positions at which the identical nucleic acid base (*e.g.*, A, T, C, G, I) occurs in both sequences to yield the number of matched positions, dividing the number of matched positions by the total number of positions in the window of comparison (*i.e.*, the window size), and multiplying the result by 100 to yield the percentage of sequence identity. For the purposes of the present invention,
30 "sequence identity" will be understood to mean the "match percentage" calculated by the DNASIS computer program (Version 2.5 for windows; available from Hitachi Software engineering Co., Ltd., South San Francisco, California, USA) using

standard defaults as used in the reference manual accompanying the software, which is incorporated herein by reference.

5 "Stringency" as used herein, refers to the temperature and ionic strength conditions, and presence or absence of certain organic solvents, during hybridisation. The higher the stringency, the higher will be the degree of complementarity between immobilised nucleotide sequences and the labelled polynucleotide sequence that remain bound following hybridization.

10 "Stringent conditions" refers to temperature and ionic conditions under which only nucleotide sequences having a high frequency of complementary bases will hybridise. The stringency required is nucleotide sequence dependent and also depends upon the various components present during hybridisation. Generally, stringent conditions are selected to be about 10 to 20°C lower than the thermal melting point (T_m) for the specific sequence at a defined ionic strength and pH. The T_m is the temperature (under defined ionic strength and pH) at which 50% of a target
15 sequence hybridises to a complementary probe.

The term "*transcribable DNA sequence*" or "*transcribed DNA sequence*", excludes the non-transcribed regulatory sequence that drives transcription. Depending on the aspect of the invention, the *transcribable* sequence may be derived in whole or in part from any source known to the art, including a
20 plant, a fungus, an animal, a bacterial genome or episome, eukaryotic, nuclear or plasmid DNA, cDNA, viral DNA or chemically synthesized DNA. A *transcribable* sequence may contain one or more modifications in either the coding or the untranslated regions which could affect the biological activity or the chemical structure of the expression product, the rate of expression or the manner of
25 expression control. Such modifications include, but are not limited to, insertions, deletions and substitutions of one or more nucleotides. The *transcribable* sequence may contain an uninterrupted coding sequence or it may include one or more introns, bound by the appropriate splice junctions. The *transcribable* sequence may also encode a fusion protein. It is contemplated that introduction into plant tissue of
30 chimeric nucleic acid constructs of the invention will include constructions wherein the *transcribable* sequence and its promoter are each derived from different species.

The term "*transformation*" means alteration of genotype by introduction of genetic material into an organism.

By "*transgenic*" is meant an organism that is transformed.

By "*transgenote*" is meant an immediate product of a transformation
5 process.

The term "*variant*" is used in the context of a nucleic acid or protein which displays a definable level of sequence identity with a reference nucleic acid or protein respectively. For example, a variant nucleic acid is hybridizable with a reference sequence under stringent conditions that are defined hereinafter, or shares a
10 percent level of sequence identity definable using a sequence comparison algorithm as hereinbefore described. Variants also encompass nucleic acids in which one or more nucleotides have been added or deleted, or replaced with different nucleotides or modified bases (eg. inosine, methylcytosine). In this regard, it is well understood in the art that certain alterations inclusive of mutations, additions, deletions and
15 substitutions can be made to a reference nucleic acid whereby the altered polynucleotide retains the biological function or activity of the reference polynucleotide. The term "*variant*" also include naturally occurring allelic variants.

By "*vector*" is meant a nucleic acid, preferably a DNA molecule derived, for example, from a plasmid, bacteriophage, or plant virus, into which a
20 nucleic acid sequence may be inserted or cloned. A vector preferably contains one or more unique restriction sites and may be capable of autonomous replication in a defined host cell including a target cell or tissue or a progenitor cell or tissue thereof, or be integratable with the genome of the defined host such that the cloned sequence is reproducible. Accordingly, the vector may be an autonomously replicating vector,
25 i.e., a vector that exists as an extrachromosomal entity, the replication of which is independent of chromosomal replication, e.g., a linear or closed circular plasmid, an extrachromosomal element, a minichromosome, or an artificial chromosome. The vector may contain any means for assuring self-replication. Alternatively, the vector may be one which, when introduced into the host cell, is integrated into the genome
30 and replicated together with the chromosome(s) into which it has been integrated. A vector system may comprise a single vector or plasmid, two or more vectors or

plasmids, which together contain the total DNA to be introduced into the genome of the host cell, or a transposon. The choice of the vector will typically depend on the compatibility of the vector with the host cell into which the vector is to be introduced. The vector may also include a selection marker such as an antibiotic
5 resistance gene that can be used for selection of suitable transformants. Examples of such resistance genes are well known to those of skill in the art.

Throughout this specification, unless the context requires otherwise, the words "comprise", "comprises" and "comprising" will be understood to imply the inclusion of a stated integer or group of integers but not the exclusion of any
10 other integer or group of integers.

2. Transcribed DNA sequences

The promoter regions of the present invention were discovered by their location adjacent to the start of DNA sequences that were found to be transcribed at high levels in one or more tissues of sugarcane (*Saccharum* sp.). A
15 hybridisation screening procedure, as described hereinafter, was used to identify genes expressed differentially in various tissues. However, it will be understood that the present invention is not restricted to use of any particular method for identifying such differentially expressed genes. For example, alternative procedures for identifying genes expressed differentially in various tissues include, but are not
20 restricted to: cDNA and genomic subtractive hybridisation as for example described by Bulman and Neill (1996, In "*Plant Gene Isolation: Principles and Practice*", G.D. Foster and D. Twell, eds Chichester, UK, Wiley, pp 369-397); multi-probe fluorescent analysis of microscopic cDNA arrays as for example described by Schena (1996 *BioEssays* 18:427-431); mRNA differential display as for example
25 described by Liang and Pardee (1992, *Science* 257:967-970) and by Callard *et al.* (1994, *BioTechniques* 16:1096-1103); computer analysis of mRNA abundance based on frequency of occurrence of identical sequences emerging from large-scale sequencing of cDNA ends (ESTs) as for example taught by Cooke *et al* (1996, *EST and genomic sequencing projects*. In *Plant Gene Isolation: Principles and Practice*,
30 *supra*, pp. 410-419); or promoter tagging by insertional mutagenesis with promoterless reporter genes as for example disclosed by Lindsey and Topping (1996,

T-DNA-mediated insertional mutagenesis. In Plant Gene Isolation: Principles and Practice, supra, pp. 275-300) and Mudge and Birch (1998, *Austral. J. Plant Physiol.* 25:637-643), which are all incorporated herein by reference.

2.1. Stem-specific transcribed sequences

5 The invention further provides DNA sequences that are transcribed preferentially at high levels in stem tissue, as compared to other tissues, of sugarcane plants. Exemplary sequences of this type are set forth in SEQ ID NO: 1, 2, 3 and 4.

10 SEQ ID NO: 1 is a cDNA of 976 nucleotides in length. This sequence has a 5' untranslated region (5' UTR) situated at position 1 to 32 and a 3' untranslated region (3' UTR) at position 594 to 881. A poly(A) tail is present at the end of this sequence. Two putative polyadenylation signals (AATAAA) are present between nucleotide 853 and nucleotide 862. An open reading frame (ORF) is present from nucleotide 33 to nucleotide 593. The nucleotide sequence and the deduced amino acid sequence (FIG. 14, SEQ ID NO: 16) do not show any homologues in the
15 databases with more than 40% overall sequence identity.

 SEQ ID NO: 2, 3 and 4 are homologous cDNA sequences (FIG. 15). The 5' UTRs are present from nucleotide position 1 to 128 (FIG. 15) and the 3' UTRs from nucleotide position 576 to the end of the sequences. The three cDNA sequences have poly(A) tails and a putative polyadenylation signal (AATAAA) is
20 present at nucleotide position 810 to 815 (FIG. 15). The longest ORF is present from nucleotide position 129 to 575 (FIG. 15). The deduced amino acid sequence (SEQ ID NO: 17) is also presented in FIG. 15. A mutation in SEQ ID NO: 3 at position 435 (FIG. 15) introduced an in-frame stop codon, but the presence of the SEQ ID NO: 3 in the cDNA library indicates that it is still being transcribed from a functional
25 promoter. Screening of the databases (nucleotide sequences and protein sequences) only revealed one similar sequence: EST Zm474 (accession number W49474) from maize. This maize EST is similar (83% sequence identity, 3 gaps) to the nucleotide sequence between nucleotide position 692 and 887 (FIG. 15), which is located within the 3' UTR.

SEQ ID NO: 9 is a partial cDNA of 597 nucleotides in length. An ORF is present from nucleotide positions 1 to 243 (FIG. 26). Its deduced amino acid sequence (SEQ ID NO: 18) is homologous to the type 2 plant metallothionein-like proteins. Examples of such proteins can be found in the nucleic acid and protein databases under accession numbers P30564 (castor bean), P43391 and P25860 (*Arabidopsis thaliana*) and others. A 3' UTR is present from nucleotide position 244 to the end of the sequence, which includes a poly(A), tail (FIG. 26).

SEQ ID NO: 10 is a cDNA of 1339 nucleotides in length. An ORF is present from nucleotide positions 124 to 1068 (FIG. 29, SEQ ID NO: 19). A 5' UTR is present from nucleotide positions 1 to 123 and a 3' UTR is present from nucleotide positions 1069 to the end of the sequence (FIG. 29). A putative polyadenylation signal is present between nucleotide positions 1301 and 1306 (TATAAA, FIG. 29). The amino acid sequence deduced from the ORF (SEQ ID NO: 19) is homologous to homeobox genes / proteins that can be found in the databases (nucleic acids and proteins) under such accession numbers as AAD13611 (maize), G4887610 (rice), BAA76750 (tobacco) and others.

SEQ ID NO: 11 is a cDNA of 1046 nucleotides in length. This sequence has a partial ORF from nucleotide positions 1 to 566 (FIG. 31). Its deduced amino acid sequence (SEQ ID NO: 20) is homologous to auxin-induced or auxin-responsive proteins found in the databases (nucleic acids and proteins) under such accession numbers as, for example, AT18409 and AT18416 (*Arabidopsis thaliana*), GMAUX28 (soybean), AF123509 and AF123508 (tobacco) and others. A 3'UTR is present from nucleotide position 567 to the end of the sequence (FIG. 31), and includes multiple putative polyadenylation signals such as AAAAAA and AATAAT, and a poly(A) tail.

SEQ ID NO: 16 is the amino acid sequence deduced from the ORF in the nucleotide sequence corresponding to SEQ ID NO: 1 (FIG. 14) (see SEQ ID NO: 1).

SEQ ID NO: 17 is the amino acid sequence deduced from the ORF in the nucleotide sequence corresponding to SEQ ID NO: 2, 3 and 4 (FIG. 15) (see SEQ ID NO: 2, 3 and 4).

SEQ ID NO: 18 is the amino acid sequence deduced from the ORF in the nucleotide sequence corresponding to SEQ ID NO: 9 (FIG. 26) (see SEQ ID NO: 9).

5 SEQ ID NO: 19 is the amino acid sequence deduced from the ORF in the nucleotide sequence corresponding to SEQ ID NO: 10 (FIG. 29) (see SEQ ID NO: 10).

SEQ ID NO: 20 is the amino acid sequence deduced from the ORF in the nucleotide sequence corresponding to SEQ ID NO: 11 (FIG. 31) (see SEQ ID NO: 11).

10 2.2. Meristem-specific transcribed sequences

SEQ ID NO: 7 is a partial cDNA of 366 nucleotides in length (FIG. 23). No homologues could be found in the nucleic acids databases.

15 SEQ ID NO: 8 is a partial sequence of a cDNA. This sequence of 395 nucleotides is homologous to cellulase (EC 3.2.1.4) nucleotide sequences such as the ones found in the nucleic acids databases under the accession numbers, for example, THFE4AA and CFICENBAA (bacteria), AF128404 (tobacco) or SLU20590 (tomato). Part of this nucleotide sequence (from nucleotide 144 to nucleotide 395, FIG. 24) can be translated into an amino acid sequence homologous to cellulase amino acid sequence (FIG. 24, SEQ ID NO: 21)

20 SEQ ID NO: 16 is the amino acid sequence derived from an ORF in the nucleotide sequence of the SEQ ID NO: 1 (FIG. 14). This amino acid sequence has no homologues in the protein databases (see SEQ ID NO: 1).

25 SEQ ID NO: 21 is the amino acid sequence deduced from the ORF in the nucleotide sequence corresponding to SEQ ID NO: 8 (FIG. 24) (see SEQ ID NO: 8).

2.3. Constitutive transcribed sequences

The invention also features DNA sequences that are transcribed constitutively in leaves, stems and roots of sugarcane. Preferred sequences of this type are set forth in SEQ ID NO: 5 and 6.

SEQ ID NO: 5 is a partial cDNA which includes the end of the coding sequence from nucleotide position 1 to 43, the 3' UTR from nucleotide position 44 to 279 and a poly(A) tail (FIG. 16). SEQ ID NO: 5 is homologous to SEQ ID NO: 6.

5 SEQ ID NO: 6 is homologous to SEQ ID NO: 5 and contains a 5' UTR from nucleotide position 1 to 91, a coding sequence from nucleotide position 92 to 349, a 3' UTR from nucleotide position 350 to 551 and a poly(A) tail (FIG. 17). The deduced amino acid sequences from SEQ ID NO: 5 and 6 (FIG. 16, SEQ ID NO: 22; FIG. 17, SEQ ID NO: 23) correspond to a ribosomal protein S27, with numerous examples of homologues found in the databases (examples can be found
10 under accession numbers D231399 for a rice mRNA sequence, X85544 for a barley mRNA or L19739 for a human mRNA).

SEQ ID NO: 22 is the amino acid sequence deduced from the ORF in the nucleotide sequence corresponding to SEQ ID NO: 5 (FIG. 16) (see SEQ ID NO: 5).

15 SEQ ID NO: 23 is the amino acid sequence deduced from the ORF in the nucleotide sequence corresponding to SEQ ID NO: 6 (FIG. 17) (see SEQ ID NO: 6).

2.4. Polynucleotide sequence variants of transcribed DNA sequences

20 In one embodiment, isolated nucleic acid variants of the invention may be prepared according to the following procedure:

- (i) obtaining a nucleic acid extract from a suitable tissue of a plant, preferably a monocotyledonous plant;
- (ii) creating primers which are optionally degenerate wherein each comprises a portion of a transcribable DNA sequence of the invention; and
25
- (iii) using said primers to amplify, via nucleic acid amplification techniques, at least one amplification product from said nucleic acid extract, wherein said amplification product corresponds to a polynucleotide sequence variant.

Suitable nucleic acid amplification techniques are well known to the skilled addressee, and include polymerase chain reaction (PCR) as for example described in Ausubel *et al.* (*supra*) which is incorporated herein by reference; strand displacement amplification (SDA) as for example described in U.S. Patent No 5,422,252 which is incorporated herein by reference; rolling circle replication (RCR) as for example described in Liu *et al.*, (1996, *J. Am. Chem. Soc.* 118:1587-1594 and International application WO 92/01813) and Lizardi *et al.*, (International Application WO 97/19193) which are incorporated herein by reference; nucleic acid sequence-based amplification (NASBA) as for example described by Sooknanan *et al.*, (1994, *Biotechniques* 17:1077-1080) which is incorporated herein by reference; and Q- β replicase amplification as for example described by Tyagi *et al.*, (1996, *Proc. Natl. Acad. Sci. USA* 93:5395-5400) which is incorporated herein by reference.

An embodiment of this method employing inverse PCR and primers according to SEQ ID NOS 24 and 25 will be described hereinafter.

Typically, polynucleotide sequence variants that are substantially complementary to a reference polynucleotide are identified by blotting techniques that include a step whereby nucleic acids are immobilised on a matrix (preferably a synthetic membrane such as nitrocellulose), followed by a hybridisation step, and a detection step. Southern blotting is used to identify a complementary DNA sequence; northern blotting is used to identify a complementary RNA sequence. Dot blotting and slot blotting can be used to identify complementary DNA/DNA, DNA/RNA or RNA/RNA polynucleotide sequences. Such techniques are well known by those skilled in the art, and have been described in Ausubel *et al.* (1994-1998, *supra*) at pages 2.9.1 through 2.9.20.

According to such methods, Southern blotting involves separating DNA molecules according to size by gel electrophoresis, transferring the size-separated DNA to a synthetic membrane, and hybridising the membrane-bound DNA to a complementary nucleotide sequence labelled radioactively, enzymatically or fluorochromatically. In dot blotting and slot blotting, DNA samples are directly applied to a synthetic membrane prior to hybridisation as above.

An alternative blotting step is used when identifying complementary

polynucleotides in a cDNA or genomic DNA library, such as through the process of plaque or colony hybridisation. A typical example of this procedure is described in Sambrook *et al.* ("Molecular Cloning. A Laboratory Manual", Cold Spring Harbour Press, 1989) Chapters 8-12.

5 Typically, the following general procedure can be used to determine hybridisation conditions. Polynucleotides are blotted/transferred to a synthetic membrane, as described above. A reference polynucleotide such as a polynucleotide of the invention is labelled as described above, and the ability of this labelled polynucleotide to hybridise with an immobilised polynucleotide is analysed.

10 A skilled addressee will recognize that a number of factors influence hybridisation. The specific activity of radioactively labelled polynucleotide sequence should typically be greater than or equal to about 10^8 dpm/mg to provide a detectable signal. A radiolabelled nucleotide sequence of specific activity 10^8 to 10^9 dpm/mg can detect approximately 0.5 pg of DNA. It is well known in the art that sufficient
15 DNA must be immobilised on the membrane to permit detection. It is desirable to have excess immobilised DNA, usually 10 μ g. Adding an inert polymer such as 10% (w/v) dextran sulfate (MW 500,000) or polyethylene glycol 6000 during hybridisation can also increase the sensitivity of hybridisation (see Ausubel *supra* at 2.10.10).

20 To achieve meaningful results from hybridisation between a polynucleotide immobilised on a membrane and a labelled polynucleotide, a sufficient amount of the labelled polynucleotide must be hybridised to the immobilized polynucleotide following washing. Washing ensures that the labelled polynucleotide is hybridised only to the immobilized polynucleotide with a desired
25 degree of complementarity to the labelled polynucleotide.

 It will be understood that polynucleotide sequence variants according to the invention will hybridise to a reference polynucleotide under at least low stringency conditions. Reference herein to low stringency conditions include and encompass from at least about 1% v/v to at least about 15% v/v formamide and from
30 at least about 1 M to at least about 2 M salt for hybridisation at 42°C, and at least about 1 M to at least about 2 M salt for washing at 42°C. Low stringency conditions

also may include 1% Bovine Serum Albumin (BSA), 1 mM EDTA, 0.5 M NaHPO₄ (pH 7.2), 7% SDS for hybridisation at 65°C, and (i) 2xSSC, 0.1% SDS; or (ii) 0.5% BSA, 1 mM EDTA, 40 mM NaHPO₄ (pH 7.2), 5% SDS for washing at room temperature.

5 Preferably, the polynucleotide sequence variants hybridise to a reference polynucleotide under at least medium stringency conditions. Medium stringency conditions include and encompass from at least about 16% v/v to at least about 30% v/v formamide and from at least about 0.5 M to at least about 0.9 M salt for hybridisation at 42°C, and at least about 0.5 M to at least about 0.9 M salt for
10 washing at 42°C. Medium stringency conditions also may include 1% Bovine Serum Albumin (BSA), 1 mM EDTA, 0.5 M NaHPO₄ (pH 7.2), 7% SDS for hybridisation at 65°C, and (i) 2 x SSC, 0.1% SDS; or (ii) 0.5% BSA, 1 mM EDTA, 40 mM NaHPO₄ (pH 7.2), 5% SDS for washing at 42°C.

 More preferably, the polynucleotide sequence variants hybridise to a
15 reference polynucleotide under high stringency conditions. High stringency conditions include and encompass from at least about 31% v/v to at least about 50% v/v formamide and from at least about 0.01 M to at least about 0.15 M salt for hybridisation at 42°C, and at least about 0.01 M to at least about 0.15 M salt for washing at 42°C. High stringency conditions also may include 1% BSA, 1 mM
20 EDTA, 0.5 M NaHPO₄ (pH 7.2), 7% SDS for hybridisation at 65°C, and (i) 0.2 x SSC, 0.1% SDS; or (ii) 0.5% BSA, 1mM EDTA, 40 mM NaHPO₄ (pH 7.2), 1% SDS for washing at a temperature in excess of 65°C.

 Other stringent conditions are well known in the art. A skilled addressee will recognize that various factors can be manipulated to optimize the
25 specificity of the hybridisation. Optimisation of the stringency of the final washes can serve to ensure a high degree of hybridisation. For detailed examples, see Ausubel *et al.*, *supra* at pages 2.10.1 to 2.10.16 and Sambrook *et al.* (1989, *supra*) at sections 1.101 to 1.104, which are incorporated herein by reference.

 While stringent washes are typically carried out at temperatures from
30 about 42°C to 68°C, one skilled in the art will appreciate that other temperatures may be suitable for stringent conditions. Maximum hybridisation typically occurs at about 20°C to 25°C below the T_m for formation of a DNA-DNA hybrid. It is well known in

the art that the T_m is the melting temperature, or temperature at which two complementary polynucleotide sequences dissociate. Methods for estimating T_m are well known in the art (see Ausubel *et al.*, *supra* at page 2.10.8).

5 In general, the T_m of a duplex DNA decreases by about 1°C with every increase of 1% in the number of mismatched base pairs.

In a preferred hybridisation procedure, a membrane (*e.g.*, a nitrocellulose membrane or a nylon membrane) containing immobilised DNA is hybridised overnight at 42°C in a hybridisation buffer (50% deionised formamide, 5xSSC, 5x Denhardt's solution (0.1% ficoll, 0.1% polyvinylpyrrolidone and 0.1%
10 bovine serum albumin), 0.1% SDS and 200 mg/mL denatured salmon sperm DNA) containing labelled probe. The membrane is then subjected to two sequential medium stringency washes (*i.e.*, 2xSSC/0.1% SDS for 15 min at 45°C, followed by 2xSSC/0.1% SDS for 15 min at 50°C), followed by two sequential high stringency washes (*i.e.*, 0.2xSSC/0.1% SDS for 12min at 55°C followed by 0.2xSSC and
15 0.1%SDS solution for 12 min).

Methods for detecting a labelled polynucleotide hybridised to an immobilised polynucleotide are well known to practitioners in the art. Such methods include autoradiography, phosphorimaging, chemiluminescent, fluorescent and colorimetric detection.

20 **3. Promoter sequences of the invention**

3.1. Promoters regions of specific transcribed DNA sequences

The invention also provides promoter regions isolated adjacent to the start of the transcribed DNA sequences described in Section 2. In particular, stem-specific and constitutive promoters for expression of chimeric or heterologous genes
25 in plants, preferably monocotyledonous plants are provided. Preferred stem-specific promoters of the invention may be selected from the polynucleotide sequences set forth in SEQ ID NO: 12 or SEQ ID NO: 13. Preferred constitutive promoters of the invention include the polynucleotides set forth in SEQ ID NO: 14 or SEQ ID NO:
15.

30 The invention also contemplates biologically-active fragments of any

one of SEQ ID NO: 12, 13, 14 or 15 as well as polynucleotide sequence variants thereof. Those of skill in the art will understand that a biologically-active fragment of a promoter sequence, when fused to a particular gene and introduced into a plant cell, causes expression of the gene at a level higher than is possible in the absence of such fragment. The activity of a promoter can be determined by methods well known in the art. For example, reference may be made to Medberry *et al.* (1992, *Plant Cell* 4:185; 1993, *The Plant J.* 3:619, incorporated herein by reference), Sambrook *et al.* (1989, *supra*) and McPherson *et al.* (U.S. Patent No. 5,164,316, incorporated herein by reference).

10 3.2. Promoter variants

Promoter variants that are substantially complementary to a reference promoter of the invention may be obtained by procedures outlined in Section 2.3.

In general, variants comprise regions that show at least 70%, more suitably at least 80%, preferably at least 90%, and most preferably at least 95% sequence identity over a reference promoter sequence of identical size ("comparison window") or when compared to an aligned sequence in which the alignment is performed by a computer homology program known in the art. What constitutes suitable variants may be determined by conventional techniques. For example, polynucleotides according to SEQ ID NO: 12, 13, 14 and 15 can be mutated using random mutagenesis (*e.g.*, transposon mutagenesis), oligonucleotide-mediated (or site-directed) mutagenesis, PCR mutagenesis and cassette mutagenesis of an earlier prepared variant or non-variant version of an isolated natural promoter according to the invention.

Oligonucleotide-mediated mutagenesis is a preferred method for preparing nucleotide substitution variants of a promoter of the invention. This technique is well known in the art as, for example, described by Adelman *et al.* (1983, *DNA* 2:183). Briefly, promoter DNA is altered by hybridising an oligonucleotide encoding the desired mutation to a template DNA, where the template is the single-stranded form of a plasmid or bacteriophage containing the unaltered or native DNA sequence of the promoter of interest. After hybridisation, a DNA polymerase is used to synthesise an entire second complementary strand of the

template that will thus incorporate the oligonucleotide primer, and will code for the selected alteration in the promoter of interest.

Generally, oligonucleotides of at least 25 nucleotides in length are used. An optimal oligonucleotide will have 12 to 15 nucleotides that are completely complementary to the template on either side of the nucleotide(s) coding for the mutation. This ensures that the oligonucleotide will hybridise properly to the single-stranded DNA template molecule.

The DNA template can be generated by those vectors that are either derived from bacteriophage M13 vectors, or those vectors that contain a single-stranded phage origin of replication as described by Viera *et al.* (1987, *Methods Enzymol.* 153:3). Thus, the DNA that is to be mutated may be inserted into one of the vectors to generate single-stranded template. Production of single-stranded template is described, for example, in Sections 4.21-4.41 of Sambrook *et al.* (1989, *supra*).

Alternatively, the single-stranded template may be generated by denaturing double-stranded plasmid (or other DNA) using standard techniques.

For alteration of the native DNA sequence, the oligonucleotide is hybridised to the single-stranded template under suitable hybridisation conditions. A DNA polymerising enzyme, usually the Klenow fragment of DNA polymerase I, is then added to synthesise the complementary strand of the template using the oligonucleotide as a primer for synthesis. A heteroduplex molecule is thus formed such that one strand of DNA encodes the mutated form of the promoter under test, and the other strand (the original template) encodes the native unaltered sequence of the promoter under test. This heteroduplex molecule is then transformed into a suitable host cell, usually a prokaryote such as *E. coli*. After the cells are grown, they are plated onto agarose plates and screened using the oligonucleotide primer having a detectable label to identify the bacterial colonies having the mutated DNA. The resultant mutated DNA fragments are then cloned into suitable expression hosts such as *E. coli* using conventional technology and clones that retain the desired promoter activity are detected. Where the clones have been derived using random mutagenesis techniques, positive clones would have to be sequenced in order to detect the

mutation.

Alternatively, linker-scanning mutagenesis of DNA may be used to introduce clusters of point mutations throughout a sequence of interest that has been cloned into a plasmid vector. For example, reference may be made to Ausubel *et al.*, *supra*, (in particular, Chapter 8.4, incorporated herein by reference) which describes a first protocol that uses complementary oligonucleotides and requires a unique restriction site adjacent to the region that is to be mutagenised. A nested series of deletion mutations is first generated in the region. A pair of complementary oligonucleotides is synthesised to fill in the gap in the sequence of interest between the linker at the deletion endpoint and the nearby restriction site. The linker sequence actually provides the desired clusters of point mutations as it is moved or "scanned" across the region by its position at the varied endpoints of the deletion mutation series. An alternate protocol is also described by Ausubel *et al.*, *supra*, which makes use of site directed mutagenesis procedures to introduce small clusters of point mutations throughout the target region. Briefly, mutations are introduced into a sequence by annealing a synthetic oligonucleotide containing one or more mismatches to the sequence of interest cloned into a single-stranded M13 vector. This template is grown in an *Escherichia coli* *dut⁻ ung⁻* strain, which allows the incorporation of uracil into the template strand. The oligonucleotide is annealed to the template and extended with T4 DNA polymerase to create a double-stranded heteroduplex. Finally, the heteroduplex is introduced into a wild-type *E. coli* strain, which will prevent replication of the template strand due to the presence of apurinic sites (generated where uracil is incorporated), thereby resulting in plaques containing only mutated DNA.

Region-specific mutagenesis and directed mutagenesis using PCR may also be employed to construct promoter variants according to the invention. In this regard, reference may be made, for example, to Ausubel *et al.*, *supra*, in particular Chapters 8.2A and 8.5, which are incorporated herein by reference.

4. Chimeric DNA constructs and expression vectors

An isolated nucleic acid promoter or variant according to the invention can be fused to a heterologous nucleic acid to form a chimeric gene. The

heterologous nucleic acid may be a foreign or endogenous DNA sequence. For the purposes of transformation and expression of the heterologous nucleic acid in plants, it is preferred that the chimeric gene includes regulatory sequences which influence expression of the heterologous nucleic acid in plants. Preferably, the chimeric gene is present in an expression vector which includes regulatory sequences that enable selective propagation in bacteria.

4.1 3' Non-translated region

A 3' non-translated sequence refers to that portion of a gene comprising a DNA segment that contains a polyadenylation signal and any other regulatory signals capable of effecting mRNA processing or gene expression. The polyadenylation signal is characterized by effecting the addition of polyadenylic acid tracts to the 3' end of the mRNA precursor. Polyadenylation signals are commonly recognized by the presence of homology to the canonical form 5' AATAAA-3' although variations are not uncommon.

The 3' non-translated regulatory DNA sequence preferably includes from about 50 to 1,000 nucleotide base pairs and contains plant transcriptional and translational termination sequences. Examples of suitable 3' non-translated sequences are the 3' transcribed non-translated regions containing a polyadenylation signal from the nopaline synthase (*nos*) gene of *Agrobacterium tumefaciens* (Bevan *et al.*, 1983, *Nucl. Acid Res.*, 11:369) and the terminator for the T7 transcript from the octopine synthase gene of *Agrobacterium tumefaciens*. Alternatively, suitable 3' non-translated sequences may be derived from plant genes such as the 3' end of the protease inhibitor I or II genes from potato or tomato, the soybean storage protein genes and the pea E9 small subunit of the ribulose-1,5-bisphosphate carboxylase (ssRUBISCO) gene, although other 3' elements known to those of skill in the art can also be employed. Alternatively, 3' non-translated regulatory sequences can be obtained *de novo* as, for example, described by An (1987, *Methods in Enzymology*, 153:292), which is incorporated herein by reference.

4.2 Optional sequences

The chimeric DNA construct of the present invention can further include enhancers, either translation or transcription enhancers, as may be required. These enhancer regions are well known to persons skilled in the art, and can include the ATG initiation codon and adjacent sequences. The initiation codon must be in phase with the reading frame of the coding sequence relating to the foreign or endogenous DNA sequence to ensure translation of the entire sequence. The translation control signals and initiation codons can be of a variety of origins, both natural and synthetic. Translational initiation regions may be provided from the source of the transcriptional initiation region, or from the foreign or endogenous DNA sequence. The sequence can also be derived from the source of the promoter selected to drive transcription, and can be specifically modified so as to increase translation of the mRNA.

Examples of transcriptional enhancers include, but are not restricted to, elements from the CaMV 35S promoter and octopine synthase genes as for example described by Last *et al.* (U.S. Patent No. 5,290,924, which is incorporated herein by reference). It is proposed that the use of an enhancer element such as the *ocs* element, and particularly multiple copies of the element, will act to increase the level of transcription from adjacent promoters when applied in the context of plant transformation.

As the DNA sequence inserted between the transcription initiation site and the start of the coding sequence, *i.e.*, the untranslated leader sequence, can influence gene expression, one can also employ a particular leader sequence. Preferred leader sequences include those that comprise sequences selected to direct optimum expression of the foreign or endogenous DNA sequence. For example, such leader sequences include a preferred consensus sequence which can increase or maintain mRNA stability and prevent inappropriate initiation of translation as for example described by Joshi (1987, *Nucl. Acid Res.*, 15:6643), which is incorporated herein by reference. However, other leader sequences, *e.g.*, the leader sequence of RTBV, have a high degree of secondary structure that is expected to decrease mRNA stability and/or decrease translation of the mRNA. Thus, leader sequences (i) that do not have a high degree of secondary structure, (ii) that have a high degree of

secondary structure where the secondary structure does not inhibit mRNA stability and/or decrease translation, or (iii) that are derived from genes that are highly expressed in plants, will be most preferred.

Regulatory elements such as the sucrose synthase intron as, for example, described by Vasil *et al.* (1989, *Plant Physiol.*, 91:5175), the Adh intron I as, for example, described by Callis *et al.* (1987, *Genes Develop.*, II), or the TMV omega element as, for example, described by Gallie *et al.* (1989, *The Plant Cell*, 1:301) can also be included where desired. Other such regulatory elements useful in the practice of the invention are known to those of skill in the art.

Additionally, targeting sequences may be employed to target a protein product of the foreign or endogenous DNA sequence to an intracellular compartment within plant cells or to the extracellular environment. For example, a DNA sequence encoding a transit or signal peptide sequence may be operably linked to a sequence encoding a desired protein such that, when translated, the transit or signal peptide can transport the protein to a particular intracellular or extracellular destination, respectively, and can then be post-translationally removed. Transit or signal peptides act by facilitating the transport of proteins through intracellular membranes, *e.g.*, vacuole, vesicle, plastid and mitochondrial membranes, whereas signal peptides direct proteins through the extracellular membrane. For example, the transit or signal peptide can direct a desired protein to a particular organelle such as a plastid (*e.g.*, a chloroplast), rather than to the cytoplasm. Thus, the chimeric DNA construct can further comprise a plastid transit peptide encoding DNA sequence operably linked between a promoter region or promoter variant according to the invention and the foreign or endogenous DNA sequence. For example, reference may be made to Heijne *et al.* (1989, *Eur. J. Biochem.*, 180:535) and Keegstra *et al.* (1989, *Ann. Rev. Plant Physiol. Plant Mol. Biol.*, 40:471), which are incorporated herein by reference.

A chimeric DNA construct can also be introduced into a vector, such as a plasmid. Plasmid vectors include additional DNA sequences that provide for easy selection, amplification, and transformation of the expression cassette in prokaryotic and eukaryotic cells, *e.g.*, pUC-derived vectors, pSK-derived vectors, pGEM-derived vectors, pSP-derived vectors, or pBS-derived vectors. Additional DNA sequences include origins of replication to provide for autonomous replication of the vector,

selectable marker genes, preferably encoding antibiotic or herbicide resistance, unique multiple cloning sites providing for multiple sites to insert DNA sequences or genes encoded in the chimeric DNA construct, and sequences that enhance transformation of prokaryotic and eukaryotic cells.

5 The vector preferably contains an element(s) that permits stable integration of the vector into the host cell genome or autonomous replication of the vector in the cell independent of the genome of the cell. The vector may be integrated into the host cell genome when introduced into a host cell. For integration, the vector may rely on the foreign or endogenous DNA sequence or any other
10 element of the vector for stable integration of the vector into the genome by homologous recombination. Alternatively, the vector may contain additional nucleic acid sequences for directing integration by homologous recombination into the genome of the host cell. The additional nucleic acid sequences enable the vector to be integrated into the host cell genome at a precise location in the chromosome. To
15 increase the likelihood of integration at a precise location, the integrational elements should preferably contain a sufficient number of nucleic acids, such as 100 to 1,500 base pairs, preferably 400 to 1,500 base pairs, and most preferably 800 to 1,500 base pairs, which are highly homologous with the corresponding target sequence to enhance the probability of homologous recombination. The integrational elements
20 may be any sequence that is homologous with the target sequence in the genome of the host cell. Furthermore, the integrational elements may be non-encoding or encoding nucleic acid sequences.

For autonomous replication, the vector may further comprise an origin of replication enabling the vector to replicate autonomously in the host cell in question.
25 Examples of bacterial origins of replication are the origins of replication of plasmids pBR322, pUC19, pACYC177, and pACYC184 permitting replication in *E. coli*, and pUB110, pE194, pTA1060, and pAM.beta.1 permitting replication in *Bacillus*. The origin of replication may be one having a mutation to make its function temperature-sensitive in a *Bacillus* cell (see, e.g., Ehrlich, 1978, *Proc. Natl. Acad. Sci. USA*
30 75:1433).

4.3 Marker genes

To facilitate identification of transformants, the chimeric DNA

construct desirably comprises a selectable or screenable marker gene as, or in addition to, the expressible foreign or endogenous DNA sequence. The actual choice of a marker is not crucial as long as it is functional (*i.e.*, selective) in combination with the plant cells of choice. The marker gene and the foreign or endogenous DNA sequence of interest do not have to be linked, since co-transformation of unlinked genes as, for example, described in U.S. Pat. No. 4,399,216 is also an efficient process in plant transformation.

Included within the terms selectable or screenable marker genes are genes that encode a "secretable marker" whose secretion can be detected as a means of identifying or selecting for transformed cells. Examples include markers that encode a secretable antigen that can be identified by antibody interaction, or secretable enzymes that can be detected by their catalytic activity. Secretable proteins include, but are not restricted to, proteins that are inserted or trapped in the cell wall (*e.g.*, proteins that include a leader sequence such as that found in the expression unit of extensin or tobacco PR-S); small, diffusible proteins detectable, *e.g.* by ELISA; and small active enzymes detectable in extracellular solution (*e.g.*, α -amylase, β -lactamase, phosphinothricin acetyltransferase).

4.3.1 Selectable markers

Examples of bacterial selectable markers are the *dal* genes from *Bacillus subtilis* or *Bacillus licheniformis*, or markers that confer antibiotic resistance such as ampicillin, kanamycin, erythromycin, chloramphenicol or tetracycline resistance. Exemplary selectable markers for selection of plant transformants include, but are not limited to, a *hyg* gene which encodes hygromycin B resistance; a neomycin phosphotransferase (*neo*) gene conferring resistance to kanamycin, paromomycin, G418 and the like as, for example, described by Potrykus *et al.* (1985, *Mol. Gen. Genet.* 199:183); a glutathione-S-transferase gene from rat liver conferring resistance to glutathione derived herbicides as, for example, described in EP-A 256 223; a glutamine synthetase gene conferring, upon overexpression, resistance to glutamine synthetase inhibitors such as phosphinothricin as, for example, described WO87/05327, an acetyl transferase gene from *Streptomyces viridochromogenes* conferring resistance to the selective agent phosphinothricin as,

for example, described in EP-A 275 957, a gene encoding a 5-enolshikimate-3-phosphate synthase (EPSPS) conferring tolerance to N-phosphonomethylglycine as, for example, described by Hinchey *et al.* (1988, *Biotech.*, 6:915), a *bar* gene conferring resistance against bialaphos as, for example, described in WO91/02071; a
5 nitrilase gene such as *bxn* from *Klebsiella ozaenae* which confers resistance to bromoxynil (Stalker *et al.*, 1988, *Science*, 242:419); a dihydrofolate reductase (DHFR) gene conferring resistance to methotrexate (Thillet *et al.*, 1988, *J. Biol. Chem.*, 263:12500); a mutant acetolactate synthase gene (ALS), which confers resistance to imidazolinone, sulfonylurea or other ALS-inhibiting chemicals (EP-A-
10 154 204); a mutated anthranilate synthase gene that confers resistance to 5-methyl tryptophan; or a dalapon dehalogenase gene that confers resistance to the herbicide.

4.3.2 Screenable markers

Preferred screenable markers include, but are not limited to, a *uidA* gene encoding a β -glucuronidase (GUS) enzyme for which various chromogenic
15 substrates are known; a β -galactosidase gene encoding an enzyme for which chromogenic substrates are known; an aequorin gene (Prasher *et al.*, 1985, *Biochem. Biophys. Res. Comm.*, 126:1259), which may be employed in calcium-sensitive bioluminescence detection; a green fluorescent protein gene (Niedz *et al.*, 1995 *Plant Cell Reports*, 14:403); a luciferase (*luc*) gene (Ow *et al.*, 1986, *Science*, 234:856),
20 which allows for bioluminescence detection; a β -lactamase gene (Sutcliffe, 1978, *Proc. Natl. Acad. Sci. USA* 75:3737), which encodes an enzyme for which various chromogenic substrates are known (*e.g.*, PADAC, a chromogenic cephalosporin); an R-locus gene, encoding a product that regulates the production of anthocyanin pigments (red color) in plant tissues (Dellaporta *et al.*, 1988, in *Chromosome*
25 *Structure and Function*, pp. 263-282); an α -amylase gene (Ikuta *et al.*, 1990, *Biotech.*, 8:241); a tyrosinase gene (Katz *et al.*, 1983, *J. Gen. Microbiol.*, 129:2703) which encodes an enzyme capable of oxidizing tyrosine to dopa and dopaquinone which in turn condenses to form the easily detectable compound melanin; or a *xylE* gene (Zukowsky *et al.*, 1983, *Proc. Natl. Acad. Sci. USA* 80:1101), which encodes a
30 catechol dioxygenase that can convert chromogenic catechols.

5. Uses of the promoters of the invention

The isolated nucleic acid promoters of the invention may be used, *inter alia*, to drive expression of a foreign or endogenous DNA sequence. Preferred agronomic properties encoded by the foreign or endogenous DNA sequence include, but are not limited to, insect resistance or tolerance, herbicide resistance or tolerance, disease resistance or tolerance (*e.g.*, resistance to viruses or fungal pathogens), stress tolerance (increased salt tolerance) and improved food content or increased yields.

The foreign or endogenous DNA sequence may comprise a region transcribed into a molecule that modulates the expression of a corresponding target gene. The molecule may be an antisense RNA or a ribozyme or other transcript aimed at downregulation of expression of the corresponding target gene.

Anti-sense regulation and the use of ribozymes and co-suppression in plants are well known in the art. However, the skilled person is referred to United States Patent 5,759,829 for an example of antisense technology and to U.S. patent 5,707,835, U.S. patent 5,747,335 and U.S. patent 5,840,874 which each provide examples of ribozyme technology. With regard to co-suppression, reference is made to U.S. patent 5,283,184, U.S. patent 5,686,649 and International Publication WO98/53083 for examples of this technology. Each of these patent documents is incorporated herein by reference.

Alternatively, the foreign or endogenous DNA sequence may encode a molecule which is readily detectable or measurable, *e.g.* β -glucuronidase or luciferase; a selectable product, *e.g.*, neomycin phosphotransferase (nptII) conferring resistance to aminoglycosidic antibiotics such as geneticin and paramomycin; a product conferring herbicide tolerance, *e.g.* glyphosate resistance or glufosinate resistance; a product affecting starch biosynthesis or modification *e.g.* starch branching enzyme, starch synthases, ADP-glucose pyrophosphorylase; a product involved in fatty acid biosynthesis, *e.g.* desaturase or hydroxylase; a product conferring insect resistance, *e.g.* crystal toxin protein of *Bacillus thuringiensis*; a product conferring viral resistance, *e.g.* viral coat protein; a product conferring fungal resistance, *e.g.* chitinase, β -1,3-glucanase or phytoalexin; a product altering sucrose metabolism, *e.g.* invertase or sucrose synthase; a gene encoding valuable

pharmaceuticals, e.g. antibiotics, secondary metabolites, pharmaceutical peptides or vaccines.

6. Introduction of isolated nucleic acids into plant cells

A number of techniques are available for the introduction of DNA into a plant host cell. There are many plant transformation techniques well known to workers in the art, and new techniques are continually becoming known. The particular choice of a transformation technology will be determined by its efficiency to transform certain plant species as well as the experience and preference of the person practising the invention with a particular methodology of choice. It will be apparent to the skilled person that the particular choice of a transformation system to introduce a chimeric DNA construct into plant cells is not essential to or a limitation of the invention, provided it achieves an acceptable level of nucleic acid transfer. Guidance in the practical implementation of transformation systems for plant improvement is provided by Birch (1997, *Annu. Rev. Plant Physiol. Plant Molec. Biol.* 48: 297-326), which is incorporated herein by reference.

In principle both dicotyledonous and monocotyledonous plants that are amenable to transformation, can be modified by introducing a chimeric DNA construct according to the invention into a recipient cell and growing a new plant that harbors and expresses the foreign or endogenous DNA sequence.

Introduction and expression of foreign or chimeric DNA sequences in dicotyledonous (broadleaved) plants such as tobacco, potato and alfalfa has been shown to be possible using the T-DNA of the tumor-inducing (Ti) plasmid of *Agrobacterium tumefaciens* (See, for example, Umbeck, U.S. Patent No. 5,004,863, and International application PCT/US93/02480). A construct of the invention may be introduced into a plant cell utilizing *A. tumefaciens* containing the Ti plasmid. In using an *A. tumefaciens* culture as a transformation vehicle, it is most advantageous to use a non-oncogenic strain of the *Agrobacterium* as the vector carrier so that normal non-oncogenic differentiation of the transformed tissues is possible. It is preferred that the *Agrobacterium* harbors a binary Ti plasmid system. Such a binary system comprises (1) a first Ti plasmid having a virulence region essential for the introduction of transfer DNA (T-DNA) into plants, and (2) a chimeric plasmid. The

chimeric plasmid contains at least one border region of the T-DNA region of a wild-type Ti plasmid flanking the nucleic acid to be transferred. Binary Ti plasmid systems have been shown effective to transform plant cells as, for example, described by De Framond (1983, *Biotechnology*, 1:262) and Hoekema *et al.* (1983, *Nature*, 303:179). Such a binary system is preferred *inter alia* because it does not require integration into the Ti plasmid in *Agrobacterium*.

Methods involving the use of *Agrobacterium* include, but are not limited to: (a) co-cultivation of *Agrobacterium* with cultured isolated protoplasts; (b) transformation of plant cells or tissues with *Agrobacterium*; or (c) transformation of seeds, apices or meristems with *Agrobacterium*.

Recently, rice, corn, pineapple and sugarcane, which are monocots, have been shown to be susceptible to transformation by *Agrobacterium*, for example as described in United States Patent No. 6,037,522, International Publication WO99/36637 and Arencibia *et al.* (1998, *Transgenic Res.* 7:213). However, some monocot crop plants have not yet been successfully transformed using *Agrobacterium*-mediated transformation. The Ti plasmid, however, may be manipulated in the future to act as a vector for these other monocot plants. Additionally, using the Ti plasmid as a model system, it may be possible to artificially construct transformation vectors for these plants. Ti plasmids might also be introduced into monocot plants by artificial methods such as microinjection, or fusion between monocot protoplasts and bacterial spheroplasts containing the T-region, which can then be integrated into the plant nuclear DNA.

In addition, gene transfer can be accomplished by *in situ* transformation by *Agrobacterium*, as described by Bechtold *et al.* (1993, *C.R. Acad. Sci. Paris*, 316:1194). This approach is based on the vacuum infiltration of a suspension of *Agrobacterium* cells.

Alternatively, nucleic acids may be introduced using root-inducing (Ri) plasmids of *Agrobacterium* as vectors.

Cauliflower mosaic virus (CaMV) may also be used as a vector for introducing of exogenous nucleic acids into plant cells (U.S. Pat. No. 4,407,956). CaMV DNA genome is inserted into a parent bacterial plasmid creating a

recombinant DNA molecule that can be propagated in bacteria. After cloning, the recombinant plasmid again may be cloned and further modified by introduction of the desired nucleic acid sequence. The modified viral portion of the recombinant plasmid is then excised from the parent bacterial plasmid, and used to inoculate the plant cells or plants.

Nucleic acids can also be introduced into plant cells by electroporation as, for example, described by Fromm *et al.* (1985, *Proc. Natl. Acad. Sci., U.S.A.*, 82:5824) and Shimamoto *et al.* (1989, *Nature* 338:274-276). In this technique, plant protoplasts are electroporated in the presence of vectors or nucleic acids containing the relevant nucleic acid sequences. Electrical impulses of high field strength reversibly permeabilise membranes allowing the introduction of nucleic acids. Electroporated plant protoplasts reform the cell wall, divide and form a plant callus.

Another method for introducing nucleic acids into a plant cell is high velocity ballistic penetration by small particles (also known as particle bombardment or microprojectile bombardment) with the nucleic acid to be introduced contained either within the matrix of small beads or particles, or on the surface thereof as, for example described by Klein *et al.* (1987, *Nature* 327:70). Although typically only a single introduction of a new nucleic acid sequence is required, this method particularly provides for multiple introductions.

Alternatively, nucleic acids can be introduced into a plant cell by contacting the plant cell using mechanical or chemical means. For example, a nucleic acid can be mechanically transferred by microinjection directly into plant cells by use of micropipettes. Alternatively, a nucleic acid may be transferred into the plant cell by using polyethylene glycol which forms a precipitation complex with genetic material that is taken up by the cell.

Also contemplated are silicon carbide or tungsten whiskers, for example as described in United States Patent No. 5,302,523.

There are a variety of methods known currently for transformation of monocotyledonous plants. Presently, preferred methods for transformation of monocots are microprojectile bombardment of explants or suspension cells, and

direct DNA uptake or electroporation as, for example, described by Shimamoto *et al.* (1989, *supra*). Transgenic maize plants have been obtained by introducing the *Streptomyces hygroscopicus bar* gene into embryogenic cells of a maize suspension culture by microprojectile bombardment (Gordon-Kamm, 1990, *Plant Cell*, 2:603-618). The introduction of genetic material into aleurone protoplasts of other monocotyledonous crops such as wheat and barley has been reported (Lee, 1989, *Plant Mol. Biol.* 13:21-30). Wheat plants have been regenerated from embryogenic suspension culture by selecting only the aged compact and nodular embryogenic callus tissues for the establishment of the embryogenic suspension cultures (Vasil, 1990, *Bio/Technol.* 8:429-434). The combination with transformation systems for these crops enables the application of the present invention to monocots. These methods may also be applied for the transformation and regeneration of dicots. Transgenic sugarcane plants have been regenerated from embryogenic callus as, for example, described by Bower *et al.* (1996, *Molecular Breeding* 2:239-249).

Alternatively, a combination of different techniques may be employed to enhance the efficiency of the transformation process, *e.g.*, bombardment with *Agrobacterium* coated microparticles (EP-A-486234) or microprojectile bombardment to induce wounding followed by co-cultivation with *Agrobacterium* (EP-A-486233).

The patent and scientific publications referred to above are all incorporated herein by reference.

7. Production and characterisation of transgenic plants

7.1 Regeneration

The methods used to regenerate transformed cells into differentiated plants are not critical to this invention, and any method suitable for a target plant can be employed. Normally, a plant cell is regenerated to obtain a whole plant following a transformation process.

Regeneration from protoplasts varies from species to species of plants, but generally a suspension of protoplasts is first made. In certain species, embryo formation can then be induced from the protoplast suspension, to the stage of

ripening and germination as natural embryos. The culture media will generally contain various amino acids and hormones, necessary for growth and regeneration. Examples of hormones utilized include auxins and cytokinins. It is sometimes advantageous to add glutamic acid and proline to the medium, especially for such

5 species as corn and alfalfa. Efficient regeneration will depend on the medium, on the genotype, and on the history of the culture. If these variables are controlled, regeneration is reproducible. Regeneration also occurs from plant callus, explants, organs or parts. Transformation can be performed in the context of organ or plant part regeneration as, for example, described in *Methods in Enzymology*, Vol. 118 and

10 Klee *et al.* (1987, *Annual Review of Plant Physiology*, 38:467), which are incorporated herein by reference. Utilizing the leaf disk-transformation-regeneration method of Horsch *et al.* (1985, *Science*, 227:1229, incorporated herein by reference), disks are cultured on selective media, followed by shoot formation in about 2-4 weeks. Shoots that develop are excised from calli and transplanted to appropriate

15 root-inducing selective medium. Rooted plantlets are transplanted to soil as soon as possible after roots appear. The plantlets can be repotted as required, until reaching maturity.

In vegetatively propagated crops, the mature transgenic plants are propagated by the taking of cuttings or by tissue culture techniques to produce

20 multiple identical plants. Selection of desirable transgenotes is made and new varieties are obtained and propagated vegetatively for commercial use.

In seed propagated crops, the mature transgenic plants can be self-crossed to produce a homozygous inbred plant. The inbred plant produces seed containing the newly introduced foreign gene(s). These seeds can be grown to

25 produce plants that would produce the selected phenotype, *e.g.*, early flowering.

Parts obtained from the regenerated plant, such as flowers, seeds, leaves, branches, fruit, and the like are included in the invention, provided that these parts comprise cells that have been transformed as described. Progeny and variants, and mutants of the regenerated plants are also included within the scope of the

30 invention, provided that these parts comprise the introduced nucleic acid sequences.

It will be appreciated that the literature describes numerous techniques for regenerating specific plant types and more are continually becoming known. Those of ordinary skill in the art can refer to the literature for details and select suitable techniques without undue experimentation.

5 **7.2. Characterization**

To confirm the presence of the heterologous nucleic acid in the regenerating plants, a variety of assays may be performed. Such assays include, for example, "molecular biological" assays well known to those of skill in the art, such as Southern and Northern blotting and PCR; a protein expressed by the heterologous
10 DNA may be analysed by western blotting, high performance liquid chromatography or ELISA (e.g., nptII) as is well known in the art.

Examples of various methods applicable to characterization of transgenic plants are provided in Chapters 9 and 11 of PLANT MOLECULAR BIOLOGY A Laboratory Manual Ed. M.S. Clark (Springer-Verlag, Heidelberg,
15 1997), which chapters are herein incorporated by reference.

So that the invention may be understood and more detail, the skilled person is directed to the following non-limiting examples.

EXAMPLE 1

Construction of a tissue-specific cDNA library

20 **Extraction of tissue-specific RNA**

Total RNA was extracted essentially according to the method of Chirgwin *et al.* (1979, Biochemistry 18 (24):5294-5299), from 9-month old Pindar sugarcane plants field-grown at the BSES Pathology Farm at Eight Mile Plains in Brisbane, Queensland, Australia. Roots, stems and leaves were harvested from three
25 separate plants randomly chosen in the field. The tissues were washed and stripped of dead material and immediately quick-frozen in liquid nitrogen. Five to ten grams of frozen tissue were ground to a fine powder in liquid nitrogen and 40 mL of a 4 M guanidine isocyanate solution were added. After thawing the mixture, 140 μ L of β -

mercaptoethanol and 2 mL of a 10% sarkosyl solution were added and thoroughly mixed. The cell debris was pelleted by centrifugation. The supernatant was then filtered through one layer of Miracloth (Calbiochem) and gently poured over a 5.7 M caesium chloride cushion. The RNA was spun down for 18 hours in a SW27 rotor at 20°C (25,000 RPM). The supernatant was carefully removed from the tubes and the RNA pellet was resuspended in 400 µL of DEPC-treated distilled water. After a phenol/chloroform extraction (phenol/chloroform/isoamyl alcohol 24/24/2), the RNA was precipitated by addition of 1/10 volume of sodium citrate (3M, pH 5.2) and 2.5 volumes of ethanol. The quantity of extracted RNA was determined by spectrophotometry (OD_{260nm}) and the quality visualised by agarose gel electrophoresis (Sambrook *et al.* 1989).

Isolation of tissue-specific mRNA

The PolyATtract® mRNA Isolation Systems (Promega) was used to separate the poly(A) RNAs from the non-poly(A) RNAs. The system uses a biotinylated oligo(dT) primer to hybridise to the poly(A) tail of the mature mRNAs. The duplexes are captured by streptavidin coupled to a paramagnetic particle and isolated using a magnetic stand. The mRNAs are then eluted directly by DEPC-treated dH₂O. The yields of mRNA from total RNA were 0.37% (leaf), 0.31% (root) and 0.27% (stem).

20 cDNA synthesis

Stem mRNAs (5µg) were used in the synthesis of double-stranded cDNAs with the TimeSaver® cDNA Synthesis Kit (Pharmacia Biotech). The first-strand cDNA was synthesized using an oligo(dT)₁₂₋₁₈ primer. After synthesis of the second strand, *EcoRI*/*NotI* adaptors were ligated to the cDNAs.

25 cDNA library

A stem-specific cDNA library was constructed in Lambda ZAP®II (Predigested Lambda ZAP®II/*EcoRI*/CIAP Cloning Kit, Stratagene). Stem cDNAs (150 ng) were ligated to 1 µg of predigested Lambda ZAP®II vector for 16 hrs at 14°C. Half of the ligation was *in vitro*-packaged (Gigapack® II Gold Packaging

Extract, Stratagene). Titration of the library was carried out using 1/500 and 1/5000 of the phage-containing supernatant and XL1-Blue MRF' host bacteria. After the phage and the host cells were incubated together, top agar containing IPTG (2.5 mM) and X-gal (4 g/L) was added and the mixture plated on NZY solid medium plates. The titer was estimated at 715,000 white plaque forming units/ μ g of vector with 4.6% non-recombinant background plaques (blue plaques). The second half of the ligations was similarly packaged. The packaged phages were combined, the titer was estimated at 663,000 pfu/mL after allowing a 20% drop of the titer after a 4-day storage at 4°C. The cDNA library was amplified by plating 50,000 pfu/ 150 mm-NZY plate (six plates). After 8hrs incubation at 37°C, when the plaques were no more than 1 to 2 mm in diameter, the plates were overlaid with 8-10 mL of SM buffer (10 mM NaCl, 8 mM MgSO₄·7H₂O, 50 mM Tris-HCl pH 7.5 and 0.01% gelatin) and gently rocked overnight at 4°C. The bacteriophage suspension was recovered from each plate and pooled. Chloroform was added (5% final concentration) and the cell debris pelleted by centrifugation (10 min at 2000xg). The supernatant was recovered and chloroform added to 0.3% final concentration. Aliquots were stored in 7% DMSO at -80°C. The titer of the amplified library was 3×10^{10} pfu/mL.

EXAMPLE 2

20 *Differential screening of the stem cDNA library, isolation of Lambda clones*

Plating of the cDNA library

The stem cDNA library was plated with *E.coli* host strain LE392, at a density of 8,000 pfu/150 mm NZY plates (supplemented with 0.2% maltose). Ten plates were incubated at 37°C for 9 hrs, when the plaques reached a size of 1 mm or less in diameter without being confluent. Plaque growth was stopped by transferring the plates to 4°C. The phage plaques were transferred onto nylon membranes (Nylon N+, Amersham) in triplicates: the first lift was in contact with the plaques for 45 seconds, the second lift for 2 min and the third lift for 4 min. The phage DNA was then denatured by placing the membrane for 2 min on a 3MM chromatography paper (Whatman) saturated with denaturing solution (2M NaOH, 1.5M NaCl). The

membrane was then transferred onto a 3MM chromatography paper saturated with a neutralising buffer (0.5 M Tris-HCl pH 8.0, 1.5 M NaCl) for 5min and finally rinsed in a 0.2 M Tris-HCl pH 8.0 and 2xSSC buffer (0.2 M NaCl, 0.03 M sodium citrate, pH 7.0) for 1min. The phage DNA was then UV-cross-linked to the damp
5 membranes (GS Gene Linker™ UV Chamber, Bio-Rad). The membranes were air-dried and stored until further use between sheets of 3MM chromatography papers.

Single-stranded radiolabelled cDNAs (probes)

Labelling reactions (30 µL total volume) contained 2 µg of mRNAs used as templates, 1 µL of Rnasin® (Promega), 6 µL of 5x buffer (250 mM Tris-HCl
10 pH 8.3, 375 mM KCL, 15 mM MgCl₂), 2 µL of dNTPs solution (10 mM each of dATP, dTTP and dGTP), 2 µL of oligo(dT) (0.5 µg/µL), 5 µL of ³²PdCTP (10 µCi/µL, Amersham) and 1 µL of M-MLV Reverse Transcriptase (Life Technologies). The mRNAs were brought to 13 µL with DEPC-treated water and denatured for 10 min at 65°C prior to the addition of the other components. The
15 reaction was carried out at 37°C for 1 hr. The probes were purified through a polyacrylamide column (Biospin 30 column, Bio-Rad).

Differential hybridisation

The nylon membranes were pre-hybridised for 6 hrs at 42°C in a pre-hybridisation solution containing 50% deionised formamide, 5xSSC (0.5 M NaCl,
20 0.075 M sodium citrate, pH 7.0), 5x Denhardt's solution (0.1% ficoll, 0.1% polyvinylpyrrolidone and 0.1% bovine serum albumin), 0.1% sodium dodecyl sulfate (SDS) and 200 mg/mL denatured salmon sperm DNA. The radiolabelled probe was added directly to the pre-hybridisation solution after completion of the pre-hybridisation procedure. Sets of membranes were hybridised as follows: the first lifts
25 were hybridised to the leaf radiolabelled cDNAs, the second lifts were hybridised to the root radiolabelled cDNAs and the third lifts were hybridised to the stem radiolabelled cDNAs.

Washing of the membranes and exposure to a X-ray film

After a 20 hr hybridisation, the membranes were washed sequentially in 2xSSC and 0.1% SDS solution for 15 min at 45°C, 2xSSC and 0.1% SDS solution for 15 min at 50°C, 0.2xSSC and 0.1% SDS solution for 12min at 55°C and 0.2xSSC and 0.1%SDS solution for 12 min at 65°C. The membranes were wrapped in a plastic film and exposed for two days to X-ray film (X-Omat Diagnostic Film, Kodak).

Primary screening

Identification of differentially expressed cDNA clones (stem-specific)

Each set of films corresponding to the same initial plate was visually examined for differentially hybridising plaques: for example, the film 1.1 (corresponding to the plate 1, first lift, probed with the leaf radiolabelled cDNAs) was overlaid with the film 1.3 (corresponding to the plate 1, third lift, probed with the stem radiolabelled cDNAs). The ten sets of films were screened for plaques hybridising to the stem radiolabelled cDNAs but giving no signals (or faint signals) when probed by either the leaf or root radiolabelled cDNAs. This method allowed the identification of 44 plaques corresponding to potentially stem-specific cDNA clones.

Identification of constitutively expressed cDNA clones

The same method revealed many plaques that strongly hybridised to each of the three different independently applied radiolabelled cDNAs (i.e., stem, root and leaf). Seven plaques were selected amongst many that fit the requirements.

Secondary screening

Identification of differentially expressed cDNA clones (stem-specific)

The plaques identified during the primary screening were cored from the plates and eluted in 1 mL of SM buffer and 20 µL of chloroform. The titers were determined for each eluate. A top agar bacterial lawn was poured onto 100 mm-NZY plates (supplemented with 0.2% maltose). About 50 phages from each eluate were

spread onto 1/4 of the plate. Plaques were grown at 37°C for 8 hrs, then placed at 4°C. Triplicate lifts were made from each plate (Nylon N+, Amersham) as described above. Pre-hybridisation, hybridisation and visual screening, carried out in the same manner as for the primary screening, confirmed 37 plaques with a possible stem-specific expression pattern.

Identification of constitutively expressed cDNA clones

Secondary screening confirmed all of seven tested constitutive clones.

In vivo excision of the pBluescript SK(-) phagemid from the Lambda ZAP II vector

The excision of the phagemid from the Lambda ZAP II vector was performed as described in the manual of the Lambda ZAP® II/EcoRI/CIAP Cloning Kit (Stratagene). The ExAssist/SOLR system procedure results in SOLR colonies containing the pBluescript SK(-) double-stranded phagemid with a cloned DNA insert. The plasmids were isolated and restricted by either *EcoRI* or *NotI* to release the cloned cDNA which was purified from an agarose gel and used to generate radiolabelled probes by random priming for Southern and northern analyses.

EXAMPLE 3

Northern and Southern analyses of the cDNA clones

Northern analysis

The specificity of the expression of each clone was verified by northern analysis. Total RNA was extracted from different tissues (i.e. roots, different stem internodes and nodes, young emerging leaves, mature leaves and older leaves) of field-grown sugarcane and maize plants and glasshouse-grown sorghum plants as described in Example 1. The RNA sample was mixed with 6 volumes of RNA sample buffer (60% deionised formamide, 25% of 37% formaldehyde and 15% of a 10xMOPS buffer containing 0.2 M MOPS pH 7.0, 50 mM sodium citrate and 5 mM EDTA pH 8.0) and 1 volume of RNA loading solution (50% glycerol, 1mM EDTA pH 8.0, 2 mg/mL bromophenol blue and 2 mg/mL xylene cyanol). Prior to loading the gel, the samples were heat-denatured at 65°C for 10 min. Equal amounts

of total RNA (8 µg/lane) were loaded onto an agarose gel. The gel was run in 1xTBE buffer (0.1 M Tris, 0.09 M Boric acid and 1 mM EDTA). Transfer of the RNA to a nylon membrane (Nylon N+, Amersham) was by capillary blotting (Sambrook *et al.* 1989). Pre-hybridisation and hybridisation were carried out as described before, using probes labelled with ³²PdCTP (Amersham Rediprime DNA labelling system). Blots were washed at the stringency previously described, and scanned with a PhosphorImager SI (Molecular Dynamics). FIGS 1 and 2 show the expression pattern of the cDNA clone c67 (SEQ ID NO: 1) that is preferentially expressed in the stem of field-grown sugarcane plants. The relative expression levels are 100 in the stem, 0.3-0.7 in the leaves and between 1.7 and 5 in the roots (depending on the cultivar). Within the stem, c67 is predominantly expressed in the more mature parts of the stem. FIGS. 3, 4, 5 and 6 show the stem-specific expression pattern of the cDNA clone c51 (SEQ ID NO: 2, 3 and 4). The c51 probe also reveals a homologous stem-specific cDNA in maize and sorghum plants (with some expression in the roots of glasshouse-grown sorghum plants). Figures 7, 8, 9 and 10 show the constitutive expression of the cDNA clone c32A (SEQ ID NO: 5 and 6) within the different parts of the sugarcane, maize, sorghum and rice plants that have been tested.

Southern analysis

DNA was extracted from sugarcane cultivars Pindar or Q117, digested by selected restriction enzymes, and electrophoresed (10 µg/lane) on an agarose gel in 1xTAE buffer (40 mM Tris-acetate, 1 mM EDTA, pH 8.3). The DNA was then denatured for 1hr (2 M NaOH, 1.5 M NaCl), neutralised for 1hr (0.5 M Tris-HCl pH8.0, 1.5 M NaCl) and equilibrated in 10xSSC prior to the transfer of the DNA onto the nylon N+ membranes (Amersham) by capillary blotting. The membranes were then treated as for the northern blots for pre-hybridisation, hybridisation to a radiolabelled probe, washes and detection by PhosphorImager (Molecular Dynamics). Southern blot analyses of the clones c67 (SEQ ID NO: 1), c51 (SEQ ID NO:2, 3 and 4) and c322 (SEQ ID NO:5 and 6) are shown respectively in FIGS. 11, 12 and 13.

EXAMPLE 4***Screening of the stem-specific cDNA library***

The stem-specific cDNA library was screened as described above, using c51 (SEQ ID NO: 2) and c32A (SEQ ID NO: 5) as probes. After the secondary
5 screening, one additional clone corresponding to c32A and two additional clones, corresponding to c51, were isolated (c322, c511 and c512, corresponding respectively to SEQ ID NOS: 6, 3 and 4).

EXAMPLE 5***Sequencing of the clones***

10 Plasmid inserts were sequenced using universal primers (T3, T7, SP6, M13 Reverse and M13 Forward) or custom-designed primers, and the ABI PRISM™ Big Dye™ Terminator Cycle Sequencing Ready Reaction Kit was used and the reactions loaded onto an ABI PRISM instrument. Sequences were obtained from both strands of the DNA.

15 The nucleotide sequence of the cDNA clone c67 (976nt) is shown in figure 14 (SEQ ID NO: 1). FIG. 15 shows the nucleotide sequence alignment of the homologous cDNA clones c51, c511 and c512 (SEQ ID NO: 2, 3 and 4). FIGS. 16 and 17 show the nucleotide sequences of the cDNA clones c32A and c322 respectively (SEQ ID NO: 5 and 6). FIGS. 23, 24, 26, 29 and 31 show the nucleotide
20 sequences of cDNA clones c19 (SEQ ID NO: 7), c3 (SEQ ID NO: 8), c18 (SEQ ID NO: 9), c53A (SEQ ID NO: 10) and c57 (SEQ ID NO: 11), respectively. In addition, FIGS. 18, 19, 20 and 21 show respectively the nucleotide sequences of the promoter sequences 67p (SEQ ID NO: 12), 32A2P2 (SEQ ID NO: 13), 32A6P2 (SEQ ID NO: 14) and 51p (SEQ ID NO: 15).

EXAMPLE 6***Promoter recovery by iPCR*****Primer design for inverse PCR reaction**

Two primers designed from the sequence of the cDNA clone c67
5 (SEQ ID NO: 1) for use in an inverse PCR reaction are 67-1 (5'
AGGACGCTTCTCAGATTGGC 3'; SEQ ID NO:24) and 67-2 (5'
GTATTCGGAGTTGCAGGTCG 3'; SEQ ID NO:25).

iPCR

Genomic DNA from sugarcane (cultivar Q117) was isolated through a
10 CsCl gradient and restricted by *SacI*. The enzyme was heat-inactivated (65°C for 10
min) and 600 ng of restricted DNA were recircularised in a total reaction volume of
400 µL containing 9 Weiss units of T4 DNA ligase (New England Biolabs). After
completion of the ligation, the reaction was extracted with phenol/chloroform and
the DNA precipitated by addition of ethanol. The PCR reaction was carried out in a
15 final volume of 50 µL comprising 150 ng of circularised DNA, 10 µL of 5x buffer
(for a final concentration of 60 mM Tris-SO₄ pH9.1, 18 mM (NH₄)₂SO₄ and 1.8mM
MgCl₂), 20 ng of primer 67-1, 20 ng of primer 67-2, 5 µL of a dNTPs solution
(2 mM each) and 1.5 µL of polymerase (ELONGASE™ Enzyme Mix, Life
Technologies). After an initial 30 s at 94°C, 35 cycles were performed consisting of
20 1 min at 94°C, 1 min at 58°C and 10 min at 68°C. A second iPCR reaction was
performed in the same conditions using 0.1 µL of the first reaction as template.
Products of the iPCR reactions were run on an agarose gel and the DNA purified
from the gel (Bresaclean, Bresatec). The purified DNA was blunt-ended using
Klenow and restricted by *SacI*. The two DNA products were cloned into pGEM®-4Z
25 (Promega) digested by *SmaI* and *SacI*. Sequencing using the T7 primer identified the
plasmid clone containing a DNA region upstream of the primer 67-1 (plasmid pG4-
67pro).

Fusion of the promoter region to a reporter gene

The full nucleotide sequence (to the ATG start codon) of the promoter region obtained by the inverse PCR method is shown in FIG. 18 (SEQ ID NO: 12). The GUS gene and *nos* terminator were excised from plasmid pBI101.3 (Clontech) as an *Xba*I-*Eco*RI fragment and ligated into the *Xba*I and *Eco*RI sites of pBluescript (Stratagene) to form plasmid pBS-GUS-3. A partial *Sac*I digest, followed by a *Bam*HI digest were carried out on the pBS-GUS-3 plasmid. Within these two sites was inserted a *Sac*I-*Bam*HI DNA fragment from the plasmid pG4-67pro to obtain plasmid p67G, with the GUS coding region translationally fused to the ATG start codon at the 3' end of the 67pro.

EXAMPLE 7

Promoter recovery by screening of a genomic library

Pindar genomic library

A Pindar sugarcane genomic library made in LambdaGEM®-11 (Promega) was plated with *E.coli* host strain KW251 on 14 NZY plates (150 mm) at a density of 24,000 pfu/plate. Duplicate lifts on nylon membranes (N+, Amersham), prehybridisation and hybridisation were carried out as for the screening of the cDNA library.

The probe was made from cDNA c322 (SEQ ID NO: 6) excised from the plasmid by an *Eco*RI-*Eco*O109I digest and randomly radiolabelled (Rediprime DNA labelling system, Amersham). Two genomic clones were isolated after a secondary screening: λ 32A2 and λ 32A6. The Lambda DNA was isolated from liquid lysates (Ausubel *et al.* 1990), restricted by different enzymes (*Pst*I, *Eco*RV, *Eco*RI, *Hind*III, *Nco*I, *Bst*XI and *Eco*O109I) and the fragments were analysed by Southern blotting. From both Lambda clones, a *Pst*I fragment containing the promoter region was cloned into pZErO-2 (Invitrogen). Sequences from the resulting plasmids (pZ-32A2P2 and pZ-32A6P2) are shown in FIGS. 19 and 20 (SEQ ID NO: 13 and 14).

H32-8560 genomic library

A sugarcane genomic library (Albert *et al.* 1992, *Plant Mol. Biol.* 20:663-671) made in Lambda EMBL4 (Stratagene) was plated with *E.coli* host strain Y1090r- on ten 150 mm-plates at a density of 35,000 pfu/plate. Duplicate lifts on
5 nylon membranes (N+, Amersham), prehybridisation and hybridisation were carried out as for the screening of the cDNA library. A *NheI* DNA fragment from the cDNA c51 (SEQ ID NO: 2) was used to generate a radiolabelled probe (Rediprime DNA labelling system, Amersham). One clone was isolated after tertiary screening (λ H51-6). The DNA fragments from an *EcoRI* and a *PstI* restriction digest corresponding to
10 hybridising signals from the clone λ H51-6 were cloned into the pZErO™-2 cloning vector (Invitrogen). These clones were designated pZ-H51-6E (*EcoRI*) and pZ-H51-6P1 (*PstI*). The pZ-H51-6E insert is 1052 bp long and contains the ATG start codon and 960 bp of sequence upstream of the ATG. The pZ-H51-6P1 insert is 3.3 kb long and contains 670 bp of sequence upstream of the ATG. The 240 bp *EcoRI-PstI*
15 fragment from pZ-H51-6E was used to re-probe the Southern blot of the λ H51-6 restriction digests. A *PstI* DNA fragment hybridising to this probe was cloned into pZErO (pZ-H51-6P2). The sequence of pZ-H51-6P2 corresponds to the sequence immediately upstream of the sequence from pZ-H51-6P1. The sequence of the promoter region of the λ H51-6 clone is shown in FIG. 21 (SEQ ID NO: 15).

20 Fusion of the promoter region to a reporter gene32A6

A primer was designed for amplification of the promoter region from pZ-32A6P2 in conjunction with the M13-Reverse primer (5' CAGGAAACAGCTATGAC 3'; SEQ ID NO:26). This primer sequence (primer
25 32A4) is homologous to the region surrounding the ATG and contains a *BamHI* restriction site at its 5' end (in bold). The sequence is 5' TAGGATCCTACCATCTTGAGATGCGG 3' (SEQ ID NO:27). The PCR reaction was carried out with a proofreading DNA polymerase from the ELONGASE Enzyme Mix (Life Technologies). The PCR product was digested by *PstI* and
30 *BamHI* and purified. The GUS gene and *nos* terminator were excised from plasmid

pBI101.1 (Clontech) as a *XbaI-EcoRI* fragment and ligated into the *XbaI* and *EcoRI* sites of the pGEM®-4Z (Promega) to form plasmid pG4-GUS-1. This plasmid was restricted by *BamHI* and *PstI* and the PCR product ligated into these two sites to form a translational fusion of the 32A6 promoter to the GUS reporter gene. This
5 plasmid construct was designated p32A6G and its fusion region confirmed by sequencing with the primer Gus-2 (5' CGCTGATCAATTCCACAG 3'; SEQ ID NO:28).

32A2

The primer 32A4 was also used to amplify the promoter region of pZ-
10 32A2P2 in conjunction with the M13-Reverse primer. The PCR reaction was carried out with a proofreading DNA polymerase from the ELONGASE Enzyme Mix (Life Technologies). The PCR product was restricted by *BamHI* and *PstI*, purified and ligated into the *BamHI* and *PstI* sites of pG4-GUS-1 to form p32A2G. The fusion region was confirmed by sequencing p32A2G with the primer Gus-2.

15 51

The 51 promoter region is present in two plasmids (pZ-H51-6P1 and pZ-H51-6P2). Fusion of the two parts of the promoter was carried out as follows: pZ-H51-6P1 was partially digested with *PstI*. The full-length linear plasmid was gel purified. The far upstream promoter region in pZ-H51-6P2 was cut from the plasmid
20 pZ-H51-6P2 as a *PstI* fragment and ligated into the *PstI* site of pZ-H51-6P1. The resulting plasmid was designated pZ-H51-6P1P2.

The GUS gene and *nos* terminator were excised from plasmid pBI101.3 (Clontech) as a *XbaI-EcoRI* fragment and ligated into the *XbaI-EcoRI* sites of the pGEM®-4Z (Promega) to form plasmid pG4-GUS-3. Two translational
25 fusions of the 51 promoter were made in pG4-GUS-3. An oligonucleotide primer (51G) homologous to the region surrounding the ATG start codon was designed to contain a *BamHI* restriction site (in bold) in its 5' end. The sequence of 51G is 5' ATGGATCCCCATTGGTGACGATCAGAAG 3' (SEQ ID NO:29). Using 51G and M13-Reverse primers, the promoter region was amplified from pZ-H51-6E with a
30 proofreading DNA polymerase from the ELONGASE Enzyme Mix (Life

Technologies). The PCR product was then restricted with *Bam*HI and ligated into the *Bam*HI site of pG4-GUS3, generating the plasmid p51SG (959bp of promoter region to the ATG). The plasmid p51SG was restricted by *Nco*I and *Sa*II and a *Sa*II – *Nco*I fragment from the plasmid pZ-H51-6P1P2 was inserted, resulting in the plasmid p51LG which contains 2133 bp of promoter region (to the ATG).

EXAMPLE 8

Functionality of the promoter: sugarcane transformation

Particle bombardment

Plasmids (GUS constructs) were isolated by rapid alkaline extraction, dissolved in TE buffer and checked for quality (intactness and absence of genomic DNA and RNA contamination) by gel electrophoresis, quantified by spectrophotometry (Sambrook *et al.* 1989), and not linearised before precipitation onto tungsten particles. Tungsten (Bio-Rad M10) was purchased in small quantities, sterilised by washing in ethanol and sterile water, and stored in water at –20°C. Tungsten stored for several years as a dry powder may become unsuitable for gene transfer.

Precipitation reactions were performed by adding the following reagents at 4°C in the listed order to a 1.5 mL microfuge tube: 5 µL selectable marker plasmid DNA (1 mg/mL pEmuKN unless specified), 5 µL non-selected plasmid DNA (1 mg/mL unless specified), 50 µL tungsten (M10, 100 mg/mL thoroughly suspended in water), 50 µL CaCl₂ (2.5 M in water), 20 µL spermidine (100 mM free base in water). The preparation was mixed immediately after addition of each reagent, with minimal delay between addition of CaCl₂ and spermidine. The tungsten was then allowed to settle for 5 min on ice, before removal of 100 µL of supernatant and resuspension of the tungsten by running the tube base across a tube rack. Suspensions were used within 15 min, at a load of 4 µL/bombardment, with resuspension of particles immediately before removal of each aliquot. Assuming the entire DNA is precipitated during the reaction, this is equivalent to 1.3 µg DNA/bombardment, on 667 µg tungsten/bombardment.

Particles were accelerated by direct entrainment in a helium gas pulse, through the constriction of a syringe filter holder into target tissues in a vacuum chamber, as described previously (Bower *et al.* 1996, *supra*).

Target tissues, selection and plant regeneration

5 Embryogenic callus cultures of sugarcane cultivar Q117 were initiated and bombarded, with osmotic treatment, followed by escape-free selection for resistance to 45 mg/L G418, and regeneration to plants as described previously (Bower *et al.* 1996, *supra*).

10 Potted plants were grown at $28 \pm 2^\circ\text{C}$ in a containment glasshouse without artificial lighting.

EXAMPLE 9

Analysis of transgenic sugarcane plants

Southern analysis

15 Regenerated plants were confirmed transgenic by Southern analysis, using the GUS coding region as a probe.

Histochemical GUS analysis

20 Histochemical detection of GUS activity was done using the substrate 5-bromo-4-chloro-3-indolyl glucuronide (X-Gluc; Jefferson 1987, *Plant Molecular Biology Reporter* 5:387-405). Sugarcane tissues (leaf segments, roots and stem slices) were immersed in assay buffer (0.05% X-Gluc, 50 mM sodium phosphate pH 7.0), vacuum infiltrated twice for 10min and incubated for 16hrs at 37°C . Following this incubation, green tissues were destained with ethanol and blue-stained tissues identified. FIG. 22 shows the GUS activity in the stem of three transgenic plants transformed with plasmid p67G. This is the first demonstrated recovery of a functional promoter from sugarcane and retention of the promoter activity and
25 specificity when re-introduced into sugarcane, which has so far shown a marked propensity to silence transgenes.

The present invention has been described in terms of particular embodiments found or proposed by the present inventors to comprise preferred modes for the practice of the invention. Those of skill in the art will appreciate that, in light of the present disclosure, numerous modifications and changes may be made in the particular embodiments exemplified without departing from the scope of the invention.

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CLAIMS

1. An isolated nucleic acid comprising a nucleotide sequence which corresponds to a promoter region of a transcribable DNA sequence that is hybridizable to a probe or primer derivable from a polynucleotide sequence selected

5 from the group consisting of: -

- (a) the polynucleotide sequence set forth in FIG. 14 [SEQ ID NO: 1];
- (b) the polynucleotide sequence set forth in FIG. 15 under designator c51 [SEQ ID NO: 2];
- 10 (c) the polynucleotide sequence set forth in FIG. 15 under designator c511 [SEQ ID NO: 3];
- (d) the polynucleotide sequence set forth in FIG. 15 under designator c512 [SEQ ID NO: 4];
- (e) the polynucleotide sequence set forth in FIG. 16 [SEQ ID NO: 5];
- (f) the polynucleotide sequence set forth in FIG. 17 [SEQ ID NO: 6];
- 15 (g) the polynucleotide sequence set forth in FIG. 23 [SEQ ID NO: 7];
- (h) the polynucleotide sequence set forth in FIG. 24 [SEQ ID NO: 8];
- (i) the polynucleotide sequence set forth in FIG. 26 [SEQ ID NO: 9];
- (j) the polynucleotide sequence set forth in FIG. 29 [SEQ ID NO: 10];
and
- 20 (k) the polynucleotide sequence set forth in FIG. 31 [SEQ ID NO: 11].

2. An isolated nucleic acid comprising a nucleotide sequence which corresponds to a promoter region of a transcribable DNA sequence selected from the group consisting of: -

- (a) the polynucleotide sequence set forth in FIG. 14 [SEQ ID NO: 1];
- 25 (b) the polynucleotide sequence set forth in FIG. 15 under designator c51 [SEQ ID NO: 2];
- (c) the polynucleotide sequence set forth in FIG. 15 under designator c511 [SEQ ID NO: 3];
- (d) the polynucleotide sequence set forth in FIG. 15 under designator c512 [SEQ ID NO: 4];
- 30 (e) the polynucleotide sequence set forth in FIG. 16 [SEQ ID NO: 5];
- (f) the polynucleotide sequence set forth in FIG. 17 [SEQ ID NO: 6];

- (g) the polynucleotide sequence set forth in FIG. 23 [SEQ ID NO: 7];
(h) the polynucleotide sequence set forth in FIG. 24 [SEQ ID NO: 8];
(i) the polynucleotide sequence set forth in FIG. 26 [SEQ ID NO: 9];
(j) the polynucleotide sequence set forth in FIG. 29 [SEQ ID NO: 10];
5 and
(k) the polynucleotide sequence set forth in FIG. 31 [SEQ ID NO: 11].
3. The isolated nucleic acid of Claim 1, wherein the nucleotide sequence is of a length in the range 100 bp to 4 kb.
4. The isolated nucleic acid of Claim 3, wherein the nucleotide sequence is of a
10 length in the range 1 kb to 4kb.
5. The isolated nucleic acid of Claim 1, wherein the isolated nucleic acid is capable of directing transcription in many or all tissues of a plant.
6. The isolated nucleic acid of Claim 1, wherein the isolated nucleic acid is capable of directing transcription preferentially in stem tissue of a plant.
- 15 7. The isolated nucleic acid of Claim 1, wherein the isolated nucleic acid is capable of directing transcription preferentially in meristem tissue of a plant.
8. The isolated nucleic acid of any one of Claims 5-7, wherein the plant is a monocotyledonous plant.
9. The isolated nucleic acid of Claim 8, wherein the monocotyledonous plant is
20 sugarcane.
10. An isolated nucleic acid which comprises a nucleotide sequence selected from the group consisting of: SEQ ID NO: 12, SEQ ID NO:13, SEQ ID NO: 14 and SEQ ID NO:15.
11. An isolated nucleic acid which comprises a biologically-active fragment of
25 any one of SEQ ID NOS: 12, 13, 14 or 15.
12. An isolated nucleic acid which has at least 60% sequence identity with any one of SEQ ID NOS: 12, 13, 14 or 15.
13. An isolated nucleic acid which is capable of hybridising to any one SEQ ID NO: 12, 13, 14 or 15 under at least low stringency conditions.
- 30 14. A chimeric gene comprising the isolated nucleic acid of any one of Claims 1, 2, 10, 11, 12 or 13 operably linked to a heterologous nucleic acid.
15. The chimeric gene of Claim 14, characterized in that said isolated nucleic

acid is capable of directing transcription of the heterologous nucleic acid in many or all tissues of a plant.

16. The chimeric gene of Claim 14, characterized in that said isolated nucleic acid is capable of directing transcription of the heterologous nucleic acid preferentially in stem tissue of a plant.
17. The chimeric gene of Claim 14, characterized in that said isolated nucleic acid is capable of directing transcription of the heterologous nucleic acid preferentially in meristem tissue of a plant.
18. An expression vector comprising the isolated nucleic acid of any one of Claims 1, 2, 10, 11, 12 or 13.
19. The expression vector of Claim 14, wherein the isolated nucleic acid comprises a nucleotide sequence selected from the group consisting of SEQ ID NOS: 12, 13, 14 or 15.
20. The expression vector of Claim 18 or 19, further comprising a heterologous nucleic acid operably linked to said isolated nucleic acid.
21. The expression vector of Claim 20, wherein the heterologous nucleic acid encodes a molecule which inhibits the expression of a target gene.
22. The expression vector of Claim 21, wherein the molecule is an antisense RNA.
23. The expression vector of Claim 21, wherein the molecule is a ribozyme.
24. The expression vector of Claim 21, wherein the molecule is capable of inhibiting expression of the target gene by co-suppression.
25. The expression vector of Claim 20, wherein the heterologous nucleic acid encodes a polypeptide.
26. A method of transforming a plant cell or tissue, comprising the step of introducing into said plant cell or tissue the isolated nucleic acid of any one of Claims 1, 2, 10, 11, 12 or 13.
27. A transformed plant cell or tissue comprising the isolated nucleic acid of any one of Claims 1, 2, 10, 11, 12 or 13.
28. A transgenic plant comprising the isolated nucleic acid of any one of Claims 1, 2, 10, 11, 12 or 13.
29. The transgenic plant according to Claim 28, wherein the transgenic plant has

an altered phenotype.

30. A transgenic monocotyledonous plant according to Claim 29.

31. A transgenic sugarcane plant according to Claim 30.

32. A progeny plant produced from the transgenic plant of Claim 28

5 33. A cell, tissue or seed obtained from the plant of any one of Claims 28-31.

34. Plasmid pG4-67pro deposited with AGAL on August 30, 1999 under accession number NM99/05995.

35. Plasmid pZ-32A2P2 deposited with AGAL on August 30, 1999 under accession number NM99/05994.

10 36. Plasmid pZ-32A6P2 deposited with AGAL on August 30, 1999 under accession number NM99/05993.

37. Plasmid pZ-H51-6P1P2 deposited with AGAL on August 30, 1999 under accession number NM99/05992.

15 38. An isolated nucleic acid comprising a polynucleotide sequence selected from the group consisting of: -

(a) the polynucleotide sequence set forth in FIG. 14 [SEQ ID NO: 1];

(b) the polynucleotide sequence set forth in FIG. 15 under designator c51 [SEQ ID NO: 2];

20 (c) the polynucleotide sequence set forth in FIG. 15 under designator c511 [SEQ ID NO: 3];

(d) the polynucleotide sequence set forth in FIG. 15 under designator c512 [SEQ ID NO: 4];

(e) the polynucleotide sequence set forth in FIG. 16 [SEQ ID NO: 5];

(f) the polynucleotide sequence set forth in FIG. 17 [SEQ ID NO: 6];

25 (g) the polynucleotide sequence set forth in FIG. 23 [SEQ ID NO: 7];

(h) the polynucleotide sequence set forth in FIG. 24 [SEQ ID NO: 8];

(i) the polynucleotide sequence set forth in FIG. 26 [SEQ ID NO: 9];

(j) the polynucleotide sequence set forth in FIG. 29 [SEQ ID NO: 10];
and

30 (k) the polynucleotide sequence set forth in FIG. 31 [SEQ ID NO: 11].

39. A polypeptide encoded by an open reading frame of the isolated nucleic acid

according to Claim 38.

40. The polypeptide of Claim 39, wherein the polypeptide comprises an amino acid sequence selected from the group consisting of:

- (i) the amino acid sequence set forth in FIG. 14 [SEQ ID NO:16];
- 5 (ii) the amino acid sequence set forth in FIG. 15 [SEQ ID NO:17];
- (iii) the amino acid sequence set forth in FIG. 26 [SEQ ID NO:18];
- (iv) the amino acid sequence set forth in FIG. 29 [SEQ ID NO:19];
- (v) the amino acid sequence set forth in FIG. 31 [SEQ ID NO:20];
- (vi) the amino acid sequence set forth in FIG. 24 [SEQ ID NO:21];
- 10 (vii) the amino acid sequence set forth in FIG. 16 [SEQ ID NO:22]; and
- (viii) the amino acid sequence set forth in FIG. 17 [SEQ ID NO:23].

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DATED this first day of September, 2000

THE UNIVERSITY OF QUEENSLAND

By their Patent Attorneys

FISHER ADAMS KELLY

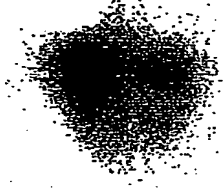
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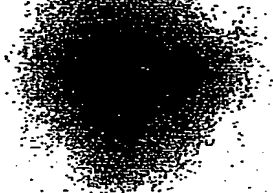
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Fig. 1

Pindar
L S R



NCo310
L S R



Q110
L S R



Q145
L S R



Fig. 2

IN₁ IN₄ IN₆ IN₈ IN₁₁ IN₁₅

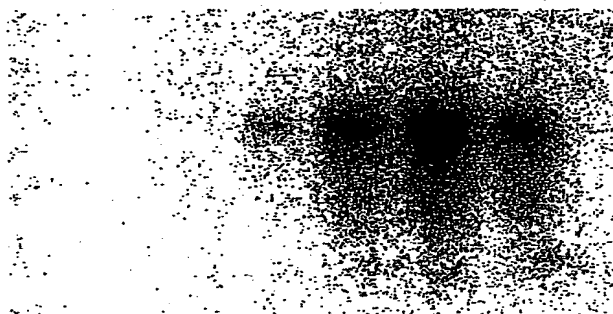


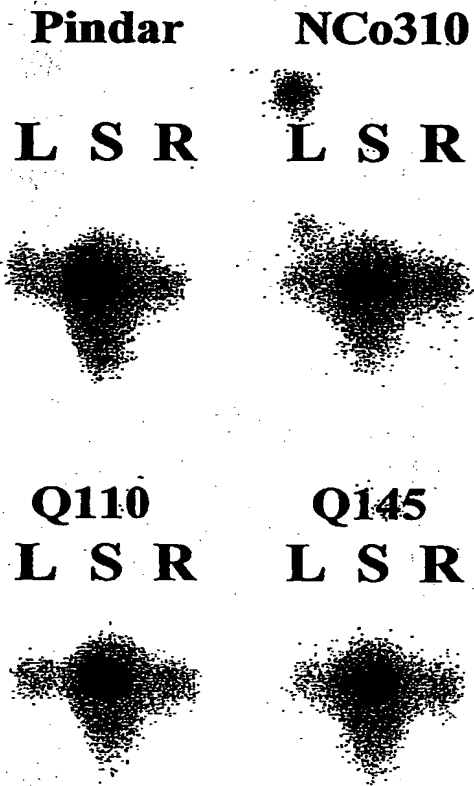
Fig. 3

Fig. 4

IN₆IN₁₁ N_c N M R YL ML OL



Fig. 5

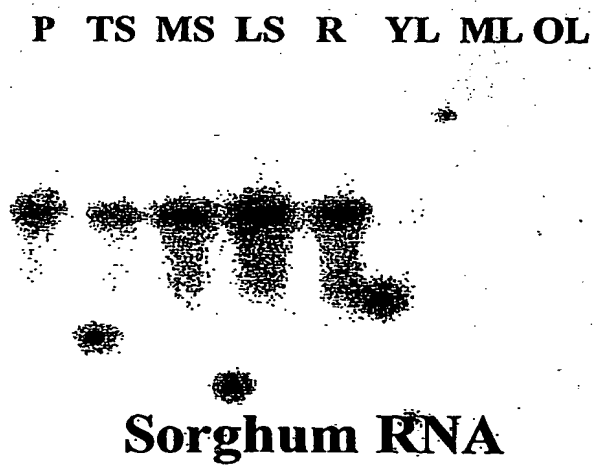
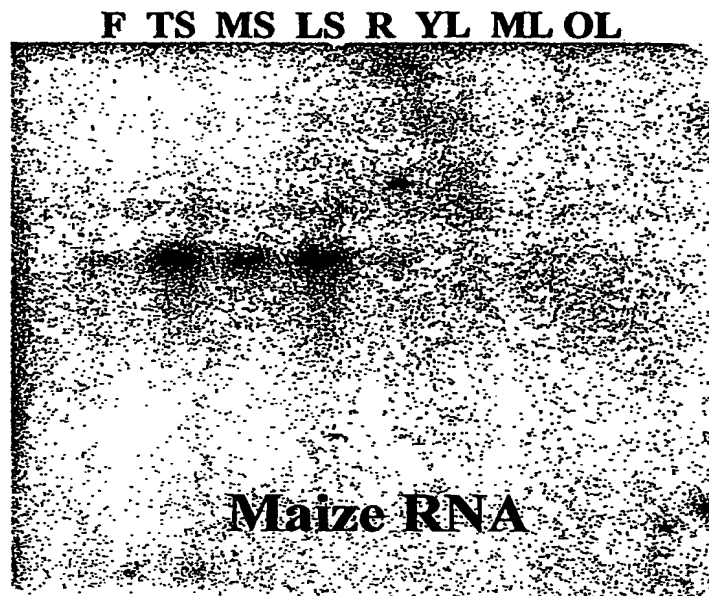


Fig. 6**Sugarcane RNA****IN₁ IN₄ IN₆ IN₈ IN₁₁ IN₁₅****Maize RNA****IN₁ IN₃ IN₅ IN₇ IN₉ IN₁₁**

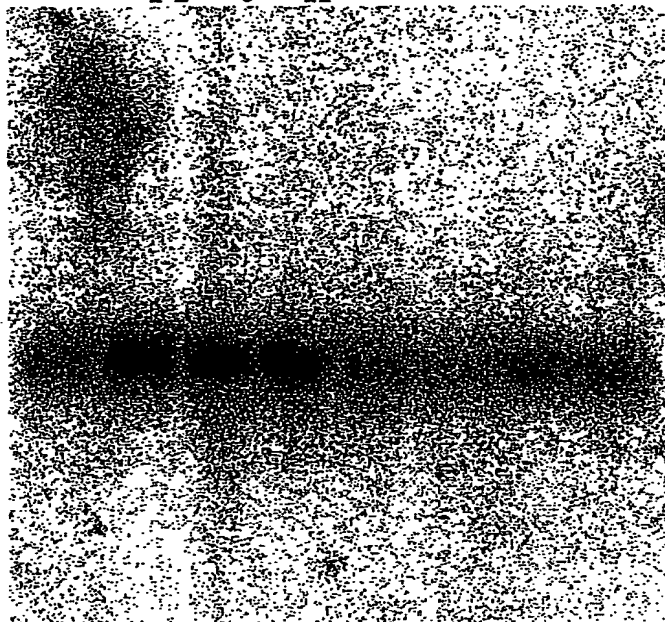
Fig. 7**N IN₁₋₂IN₆IN₁₁ R YL ML OL**

Fig. 8

Pindar

NCo310

L S R

L S R



Q110

Q145

L S R

L S R



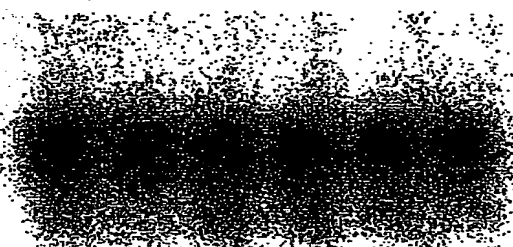
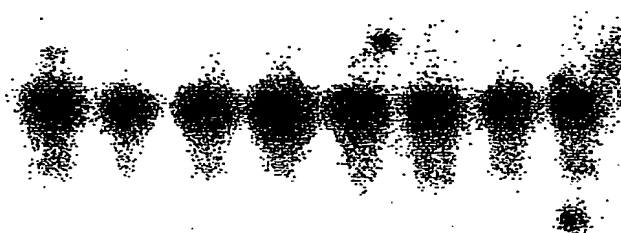
Fig. 9**Sugarcane RNA****IN₁ IN₄ IN₆ IN₈ IN₁₁ IN₁₅****Maize RNA****IN₁ IN₃ IN₅ IN₇ IN₉ IN₁₁**

Fig. 10

Sorghum RNA

P TS MS LS R YL ML OL



Rice RNA

S₁ S₂ R L₁ L₂ L₃



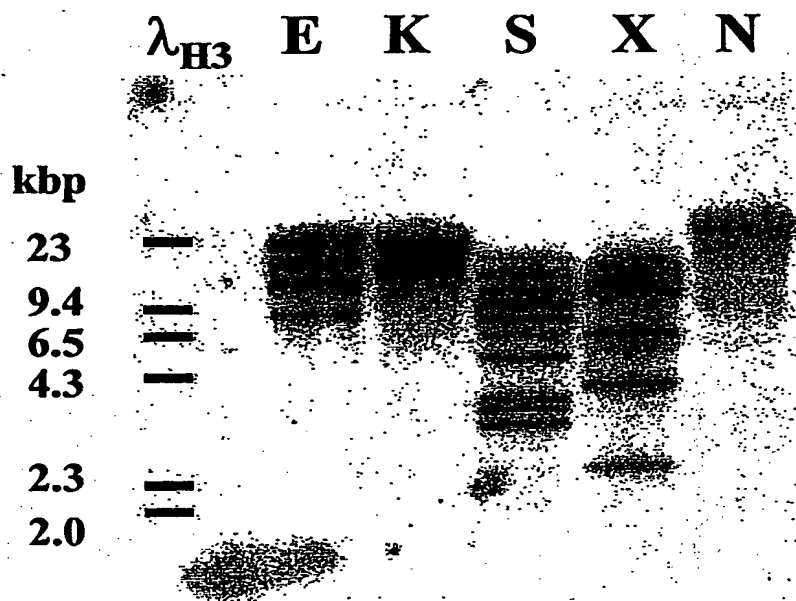
Fig. 11

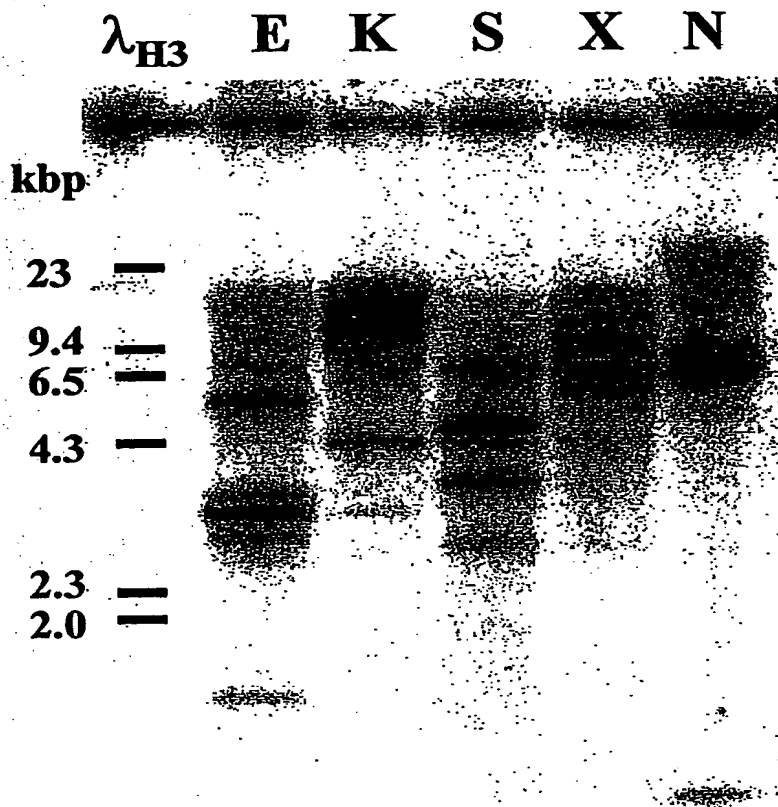
Fig. 12

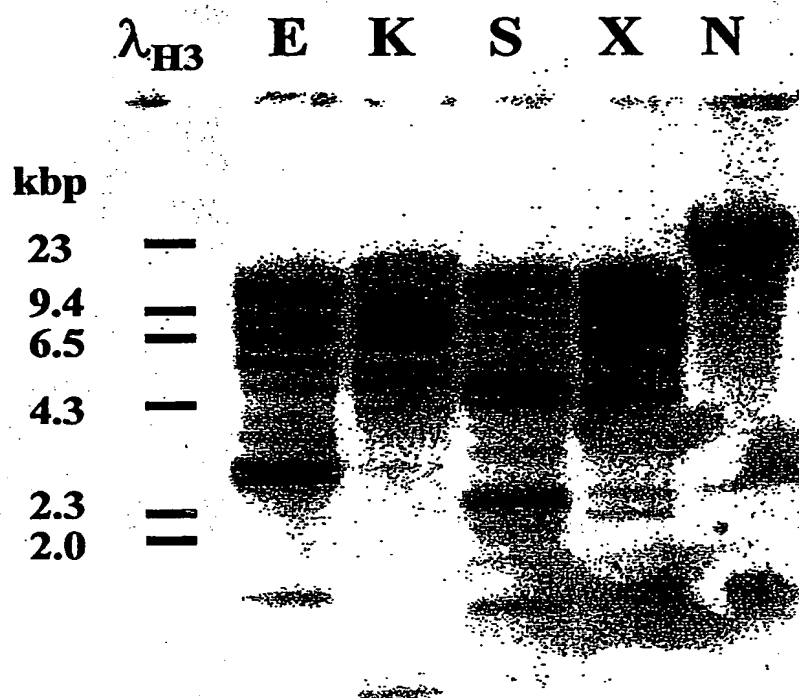
Fig. 13

Fig. 14

1 TGAAATAAAC GGTAGCTGCC ATAAC TAGTA CAATGGCCAA TCTGAGAAGC 50
M A N L R S
51 GTCCTAGCTG TGAGCCTAGC CGTGGCACTT TTCGCAGTTG CTCCTGCATC 100
V L A V S L A V A L F A V A P A S
101 GTTCGCACTG GATGAGAAAG AGTTGCACCT GAGTTTGTAC TTAAACCAGA 150
F A L D E K E L H L S L Y L N Q T
151 CATA CAGCGG AAACGGCCTT AACCAGGCGG TGGTGGTCGA ACCAGGCCTA 200
Y S G N G L N Q A V V V E P G L
201 CCTGGGGAGT TCGGCAACAT CGCCGTCCAG GACTGGCCTG TGACCAATGG 250
P G E F G N I A V Q D W P V T N G
251 GGAAGGTAGC GACGCAACCG TCGTTGGACG TGCACAGGGC ATCCAGTTCA 300
E G S D A T V V G R A Q G I Q F K
301 AACCAAGCCA GAGGAACGAC CAAGCCTGGT ATACCACCTT GACCATAGTG 350
P S Q R N D Q A W Y T T L T I V
351 TTCGAGAACA CGAGCCTCAA GGGATCCACG CTTCAGATGA TGGGTTACAT 400
F E N T S L K G S T L Q M M G Y I
401 CCCACAAGAT GGTCA GTGGA GCATTTTTGG AGGAACTGGA CAACTTACGA 450
P Q D G Q W S I F G G T G Q L T M
451 TGGCAGCGGG TGTGTGAAC CACAAGGTTG TGCGCCAAAC CAATGGCGGG 500
A R G V V N H K V V R Q T N G G
501 AGGATGTATA AGATCAACAT ACATGCCTTC TATACCCCCC TGGGCGCTTC 550
R M Y K I N I H A F Y T P L G A S
551 TAGCAACTGT GGGATTAACC TTAGGCGCTT GGACTTCGAC GCTTGATCGA 600
S N C G I N L R R L D F D A *
601 CTAGCGCGGA CTACAACAGG AGGACCGTGT TCTTCGTCGA CGCTTAATGC 650
651 ATGGAAACTT CCCACGGGGG ACCGTGTTCT TCATGGACGC TAGACCAACC 700
701 ATAATTTCTT TTCCGTTTGT ACTGTCAACA AATATAAATA TGTAAGCAT 750
751 AAATCCGAAC TGTATTCGGA GTTGCAGGTC GTCGTTGCCC CTGCCTTATG 800
801 GCCTGCACTG TACATGTACA TGTTTCTGTC AAGTTCTGCG AGTATTTTAA 850
851 GTAATAAATA AAGTG GTTGG TTTCACGGTT TAAAAAAAAA AAAAAAAAAA 900
901 AAAAAAAAAA AAAAAAAAAA AAAAAAAAAA AAAAAAAAAA AAAAAAAAAA 950
951 AAAAAAAAAA AAAAAAAAAA AAAAAA 976

Fig. 15

	1				50
c51	TCGAATGCTC	GATCGATCCC	ACTCTCAGCT	GATCGCTCAC	TCTTGCAAGCT
c511TGCTC	GATCGATCCC	ACTCTCAGCT	GATCGCTCAC	TCTTGCAAGCT
c512TCTC	GATCGATCCC	ACTCTCAGCT	GATCGCTCAC	TCTTGCAAGCT
	51				100
c51	CGATCAGTCT	TAGCTCTAGC	CTCTAGCTAG	CCAACTAGCC	GCTCCTTCGT
c511	CGATCAGTCT	TAGCTCTAGC	CTCTAGCTAG	CCAACTAGCC	ACTCCTTCGT
c512	CGATCAGTCT	TAGCTCTAGC	CTCTAGCTAG	CCAACTAGCC	ACTCCTTCGT
	101				150
c51	GTAGCCATCA	GCCTTCTGAT	CGTCACCAAT	GGCCACTGCC	GAGGTCCAGA
c511	GTAGCCATCA	GCCTTCTGAT	CGTCACCAAT	GGCCACTGCC	GAGGTCCAGA
c512	GTAGCCATCA	GCCTTCTGAT	CGTCACCAAT	GGCCACTGCC	GAGGTCCAGA
aa			M	A T A	E V Q T
	151				200
c51	CCCCGACCGT	CGTGGCGACC	GAGGAGGCGC	CCGTGGTGGG	GACGCCGGCG
c511	CCCCGACCGT	CGTGGCGACC	GAGGAGGCGC	CCGTGGTGGG	GACGCCGGCG
c512	CCCCGACCGT	CGTGGCGACC	GAGGAGGCGC	CCGTGGTGGG	GACGCCGGCG
aa	P T V	V A T	E E A P	V V E	T P A
	201				250
c51	CCGGCCGTCG	TGCCCAGAGG	GGCTGCCCCC	GCCCCCGCCG	AGGCTGAGAC
c511	CCGGCCGTCG	TGCCCAGAGG	GGCTGCCCCC	GCCCCCGCCG	AGGCTGAGCC
c512	CCGGCCGTCG	TGCCCAGAGG	GGCTGCCCCC	GCCCCCGCCG	AGGCTGAGCC
aa	P A V V	P E E	A A P	A P A E	A E P
	251				300
c51	GGCGGCCGTCG	CCCGAGGAGG	CTGCCCCCGC	CGAGGCCAAG	GTGGAGGAGC
c511	GGCGGCCGTCG	CCCGAGGAGG	CTGCCCCCGC	CGAGGCCAAG	GTGGAGGAGC
c512	GGCGGCCGTCG	CCCGAGGAGG	CTGCCCCCGC	CGAGGCCAAG	GTGGAGGAGC
aa	A A V	P E E A	A P A	E A K	V E E P
	301				350
c51	CTGCCGCCCC	GGCGGAGCCT	GAGCCTGCCG	CCGCTGAGCC	CGAGGCCGAG
c511	CTGCCGCCCC	GGCGGAGCCT	GAACCTGCCG	CCGCTGAGCC	CGAGGCCGAG
c512	CTGCCGCCCC	GGCGGAGCCT	GAGCCTGCCG	CCGCTGAGCC	CGAGGCCGAG
aa	A A P	A E P	E P A A	A E P	E A E
	351				400
c51	CCTGCCGCGG	CGGAGCCGGA	GGCGGCCCT	GCAGCCGCGG	CGGAGGAAGA
c511	CCTGCCGCGG	CGGAGCCGGA	GGCGGCCCT	GCAGCCGCGG	CGGAGGAAGA
c512	CCTGCCGCGG	CGGAGCCGGA	GGCGGCCCT	GCAGCCGCGG	CGGAGGAAGA
aa	P A A A	E P E	A A P	A A A A	E E E
	401				450
c51	GGCGCCAAAG	GAGGCGGAGC	CGGCGGCCGT	TGAGGAGGTG	AAGGAGGAGG
c511	GGCGCCAAAG	GAGGCGGAGC	CGGCGGCCGT	TGAGTGGGTG	AAGGAGGAGG
c512	GGTGCCTAAG	GAGGCGGAGC	CGGCGGCCGT	TGAGGAGGTG	AAGGAGGAGG
aa	A(v)P K	E A E P	A A V	E E V	K E E E
	451				500
c51	AGGCGGCCGC	GCCCCGTGCC	GAGACAGAGC	CGGCGGCCGC	CGAGCCCGAG
c511	AGGCGGCCGC	GCCCCGTGCC	GAGACAGAGC	CGGCGGCCGC	CGAGCCCGAG
c512	AGGCGGCCGC	GCCCCGTGCC	GAGACAGAGC	CGGCGGCCGC	CGAGCCCGAG
aa	A A A	P A A	E T E P	A A A	E P E

Fig.15 cont'd

	501				550
c51	GTTGCTGCTC	CTGCTGCCTC	CGAGGAGCCC	ACCGCGGCCG	AGCCGGCCCG
c511	GCTGCTGCTC	CTGCTTCCTC	CGAGGAACCC	GCCGCGGCCG	AGCCGGCCCG
c512	GCTGCTGCTC	CTGCTGCCTC	CGAGGAGCCC	ACCGCGGCCG	AGCCGGCCCG
aa	V(A)A	A P	A A S	E E P	T A A E P A A
	551				600
c51	CGCGGAGCCC	GAGAAGGCCA	GCGAGTGAGG	CCGTGCGCGC	GCGCAGCGGC
c511	CGCGGAGCCC	GAGAAGGCCA	GCGAGTGAGG	CCGT..GCGC	GCGCAGCGGC
c512	CGCGGAGCCC	GAGAAGGCCA	GCGAGTGAGG	CCGTGCGCGC	GCGCAGCGGC
aa	A E P	E K A S	E *		
	601				650
c51	GGCGGCCAGG	GGATCGGAGT	GAGATGGCTC	ATCGTGCGCG	CGTGACGTAG
c511	GGCGGCCAGG	GGATCGGAGT	GAGGTGGCTC	ATCGTGCGCG	CGTGACGTAG
c512	GGCGGCCAGG	GGATCGGAGT	GAGATGGCTC	ATCGTGCGCG	CGTGACGTAG
	651				700
c51	CGGCGTGGCA	GTCGCGTGCG	CGCGTACATG	GCACGGTCTT	TTGCTGTTCA
c511	CGGCGTGGCA	GTCGCGTTTG	CGCGTACATG	GCACGGTCTT	TTGCTGTTCA
c512	CGGCGTGGCA	GTCGCGTGCG	CGCGTACATG	GCACGGTCTT	TTGCTGTTCA
	701				750
c51	GTACGCTACG	GTTACTGGCG	TTTC..TGTG	TGTGTGTCGT	AGCTCGGTAC
c511	GTACGCTACG	GTTACTGGCG	TTTCTGTGTG	TGTGTGTCGT	AGCTCGGTAC
c512	GTACGCTACG	GTTACTGGCG	TTTC..TGTG	TGTGTGTCGT	AGCTCGGTAC
	751				800
c51	GTAGCTAGCA	GTGGCTGTCTG	CGTGGACTGT	CCGGTGTGGA	CTGCGAGTGA
c511	GTAGCTAGCA	GTGGCTGTCTG	CGTGGACTGT	CCGGTGAGGA	CTGCGAGTGA
c512	GTAGCTAGCA	GTGGCTGTCTG	CGTGGACTGT	CCGGTGTGGA	CTGCGAGTGA
	801				850
c51	CGTGGGCTAA	ATAAAGTGTG	CGTCGTGTGG	TGG...TGGT	GGTGGGGCCC
c511	CGTGGGCTAA	ATAAAGTGTG	CGTGGTGGTG	GGGCCCCTG	TCAGTGGCGC
c512	CGTGGGCTAA	ATAAAGTGTG	CGTCGTGTGG	TGG...TGGT	GGTGGGGCCC
	851				900
c51	GGTGCCCCGT	GCCCTGCCGC	TGCAGCTAGT	CTGTCTTTT.	.GTGTTTGT
c511	GGTGCCCCAT	GCCCTGCCGC	TGCAGCTTGT	CTGTCTTTT.	GTGTCGTGT
c512	GGTGCCCCGT	GCCCTGCCGC	TGCAGCCTGT	CTGTCTTTT	GTGTTGTGT
	901				950
c51	GGTTCCTGTT	CTCTGTCGAT	CTATATTTAT	CCTCATGTTT	GTATGGCAGT
c511	GGTTCCTGTT	CTCTGTCGAT	CTATATTTAT	CCTCATGTTT	GTATGGCAGT
c512	GGTTCCTGTT	CTCTGTCGAT	CTATATTTAT	CCTCCA ₃₁
	951			979	
c51	TTGGGTATGG	GTATCA ₄₉		
c511	TTGGGTATGG	GTATCAAACA	TGTTTGCGA ₃₀		
c512		

Fig.16

1	TGCCAGGCTC	ACCGAGGGGT	GCTCCTTCCG	TCGCAAGGGC	GATTAGATCC	50									
	A	R	L	T	E	G	C	S	F	R	R	K	G	D	*
51	TGCTGTCTTC	ATGGGAGAAG	AGAGAAATTT	GCTGTTTTAC	TACCTTCCCA	100									
101	TGATATGTAC	TCGTTGAGGA	TTTTGTTGAT	TATTATGGCT	GTTTAGCGTG	150									
151	CCCTGGCAAT	GCTTTTGTAA	ACGTGCACTT	TGCTTGAGCT	TAGTGACATC	200									
201	TACTAAGGTG	CTGTTTGGTT	TTGCTAAGGG	AGTGGCAATG	GTAATGAAAT	250									
251	CAGTTGCTGG	CGTTAATTGT	TGGGGTTTTA	AAAAAAAAAA	AAAAAAAAAA	300									
301	AAAAAAAAAA	AAAAAAAAAA	AAAAAAAAAA	AAAAAAAAAA	AAAAAAAAAA	350									
351	AAAAAAAAAA	AAAAAAAAAA	AAAAAAA	378											

Fig.17

1	GCCGCCGCTA	CGCGCTCGCC	TTTGCTCCCT	CTGTTTCCCT	TCCTCCATAC	50
51	AGGCGCAGGC	CAGGAACGCA	CAAGGCGACC	GCATCTCCAA	GATGGTGCTG	100
				M V L		
101	CAGAACGACA	TCGACTTGCT	CAACCCGCCG	GCAGAGCTTG	AGAAGCTAAA	150
	Q N D I	D L L	N P P	A E L E	K L K	
151	GCACAAGAAG	AAGCGCCTCG	TCCAGTCCCC	CAACTCCTTC	TTCATGGATG	200
	H K K	K R L V	Q S P	N S F	F M D V	
201	TCAAGTGCCA	GGGCTGTTTC	AGCATAACCA	CTGTGTTCAG	CCACTCCCAG	250
	K C Q	G C F	S I T T	V F S	H S Q	
251	ACTGTGGTTG	TGTGCCCAGG	CTGCCAAACT	GTTCTCTGCC	AACCTACTGG	300
	T V V V	C P G	C Q T	V L C Q	P T G	
301	TGGGAAGGCC	AGGCTCACCG	AGGGGTGCTC	CTTCCGTCGC	AAGGGCGATT	350
	G K A	R L T E	G C S	F R R	K G D *	
351	AGATCCTGCT	GCCTTTTAAT	GGGAGAAGAG	AGAAATTTGC	TGTTTTACAA	400
401	CCTTCCCATG	ATATGTACTC	GTTGAGGATT	TTGTTAATTA	TTATGGCTGT	450
451	TTAGCTTGCC	CTGTCAATGC	TTTTGTAAAC	GTGCACTTTG	CTTGGTGCTG	500
501	TTTGGTTTTG	CCAAGGGATT	GGCAATGGTA	GTGAAATCAG	TTGCTGACGT	550
551	TAAAAAAAAA	AAAAAA	567			

Fig. 18

1	GAGCTCTCAA	TGGGAGGTGC	TCGAAGACAT	ATTACCCAAG	TGTATGGCAA	50
51	GATGTTTAGC	TAGTAACTGA	CTGATAGTGT	AAACGATCTC	CAATGGGGCA	100
101	AGACATATTA	CCTAAGGCCA	GGCTGGTTTT	TGCAAGTTCG	AGTAGGATAT	150
151	AGAGATTCTC	GTGCGAGTTG	TAAACGATCT	CCAATGGGGC	AAGACATCCT	200
201	ACCCTATATA	TAGTGAAGGG	GCAGTAGCTG	ATTGAGAATC	AATCAATCAA	250
251	GCACAATATA	ATTTATTAAT	TTTTTATTCA	AACCCAATTT	TTTCCTTTT	300
301	CCAACCCTAA	TTATAGTTCT	CCTTTTGCCT	CTAGGACAAA	TTGACGTGTT	350
351	CCGGGTATCC	TGCTGAATCA	AGAACAACCC	TAGGTGCACC	TGCCCCGATA	400
401	GAGTCCCACC	TGGGTAGGCA	TTCATAGGGA	TTCGGGTATT	TCCTGCAAAA	450
451	AAGCGATTAA	GCTGGCTTCT	AAAACCTGGT	AGGCCGGATT	CTGTGGCCTT	500
501	CACTACCAGG	TGATTTTCAT	GTGATCCGTG	CATTCTAGCA	CTTGCTATG	550
551	TAACCCAAAC	TGAAGTCGAC	AACTATAAAT	ATGCTACTTG	CAGGATGTTA	600
601	TCACGACACA	ACTCCCAGTC	TACGAAGCCT	AAGTTTAGTT	TTGCTCGGAG	650
651	ACAAGCAATT	GTGGCCAGTC	ACTATAGCTA	CGTCAGAGGG	TAGTGGGAGC	700
701	AGTTGCGTCG	TTGGATTGAA	AACAGGTGGA	TCGTATCAGA	TATTATGCAT	750
751	TCACATGAAC	AGTAAATGTG	GTACAGTACT	TCGCAAACAA	TAAAATCTGT	800
801	CACAATTTAT	TAGTGCACTC	CTGTGACGTA	AATGCTTCTA	CGTCAGAGGA	850
851	TTTGAGTCCG	AGGGCTGCTG	CACCCATCAC	TAATGACGGT	CTTTACCCAT	900
901	CATCATGGAC	CATTGTTCAC	ATCCATGCTA	TCACTGTCGT	CCTGTCCATG	950
951	CACTGCAGCC	CTCTATAAAT	ACTGGCACCC	CTCCCCCGTT	CACAGATCAC	1000
1001	ACCACACAAG	CAAGAAATAA	ACGGTAGCTG	CCATAACTAG	TACAATG	1047

Fig. 19

1	CTGCAGAACG	ACATCGACTT	GCTCAACCCG	CCGGCAGAGC	TTGAGAAGCT	50
51	AAAGCACAAG	AAGAAGCGCC	TCGTCCAGTC	CCCCAACTCC	TTCTTCATGG	100
101	TATACCTCTT	CATGCCCTTT	TCTAATTCTG	CGTGATGGGG	TGTTGCGAAT	150
151	CATGCCCTAA	AGAGACCCAA	CGAATGTATC	ATTGGAATTT	AGTTTTGGCA	200
201	AAATAAAGAC	CTGCACCTAA	TAGGGTGTAT	TGGTTTGGAA	GAGAGTCCAG	250
251	AAATTAGATT	TAGTATCGTT	AACTACATCA	AAAATTACTT	CCAACAAAGC	300
301	AACAAGACAG	CTTAAGTGCT	CTTGTTTGCC	ATGTAACATA	CATGTAGTCA	350
351	AGCTGGATGC	TATAGGTTAT	GCATTTTTCA	TCAAATAAGT	TGTGTTTATA	400
401	AGAGAGCTAA	AATGTGAAAA	CAGAAATGAA	AATGAATCTT	AAAGTTACAA	450
451	TGTATCGACT	TATATTCATA	TTAATTAATA	TTAATATTAG	TCGTGTCTGC	500
501	TCTTGTGCCT	GATCGGAGAC	TGACTCTAAT	CATGTTATTT	TTCGTTTTTT	550
551	TTCTCCTTTG	TGTGCAGGAT	GTCAAGTGCC	AGGGCTGTTT	CAGCATGTAA	600
601	GTGTTTATGA	TTCTTGCTCC	TTTTTTGTTT	TTAGTTACTA	CAGTTGCAAT	650
651	TGATATAGAC	GCCGTGTCAT	TCCATTTATC	CATTGCTGAT	TGCTGATGTG	700
701	ATCTCCTAAT	CTTGTGTTAC	TGGTTTTCCA	TTGACAGTTT	AGCATGATCA	750
751	GTTTACTAGT	AGGGTTGCGA	TTACTACTAG	CTAGTTAAAA	TATGAAAATT	800
801	CTTGGTTTAG	TTATAAGGTT	ATATATTGAT	TTCGTAAAAAT	TTCATACCCC	850
851	CTCACTTCCC	GAATATGTAG	GACAACAAAA	TTGTTTATAG	ATGGAGATGG	900
901	AAACTGTTAA	TGTTTGACTC	CTGTTTTTGT	TTCTTTCTCA	CCTGACCAGT	950
951	ACAAGTACAA	TTGTTCTGTT	TAAATGTGGG	TTAATTTGGA	TTCAACAACA	1000
1001	ACAACAACAA	CAACATAGCC	TTTTGTCCCA	AGCAAGTTGG	GGTAGGCTAG	1050
1051	AGATGAAACC	CAAAAGAGAC	GAGAAACAGG	GAGACACAAC	GTTACACCTC	1100
1101	GCTGCTTTAT	TTGGATTACG	CTTGTTATTC	TTTTGATAAT	TGCACAACGT	1150
1151	TACACCTAGC	TGCTTTATTT	GCTGCTTGCA	AGTGTAGCTG	GTTTGATCAT	1200
1201	GTTGTGTACG	AGTTCTAATT	TACGTGTGAC	CACTAAGCTT	TACTGCAATT	1250
1251	ATCTGCGATC	TTTAAATGTT	ATCCTTTGTT	GGAAGGTTAT	TATGGTTGTA	1300
1301	TAGCTTCTGT	GATATCACGA	TTCTGAACAA	ATCAAATGTT	TGCTGTAGCT	1350
1351	TACAAAGTTT	TTGTGATTGC	AGAACCACTG	TGTTTCAGCCA	CTCCCAGACT	1400
1401	GTGGTTGTGT	GCCCAGGCTG	CCAAACTGTT	CTCTGCCAAC	CTACTGGTGG	1450
1451	GAAGGCCAGG	CTCACCAGAG	GGTGCTCCTT	CCGTCGCAAG	GGCGATTAGA	1500
1501	TCCTGCTGTC	TTTTAATGGG	AGAAGAGAGA	AATTTGCTGT	TTTACAACCT	1550
1551	TCCCATGATA	TGTACTCGTT	GAGGATTTTG	TTAATTATTA	TGACTGTTTA	1600
1601	GCTTGCCCTG	TCAATGCTTT	TGTAAACGTG	CACTTTCCTT	GAGCTTAGTG	1650
1651	ACATCTACTT	AAGGTGCTGT	TTGGTTTTGC	CAAGGGATTG	GCAGTGGTAG	1700
1701	TGAAATCAGT	TGCTGACGTT	ATTTGTTGGG	GTTTAACTGA	AAGTGTCAAT	1750
1751	TGTTTAGTTG	CCTTGTGTAA	TGGATTCAAT	TTAACAACCTG	ACTAGTACGG	1800
1801	TCTCTAGCCC	AATGCAGCGT	TCACTCTCAG	CGGCAGATTG	TTCTGTAGCA	1850
1851	GTTAACGGTC	ATAGGAGTGC	CCTCGCGCTC	GCCTTTGTCC	CCTCTCTCCC	1900
1901	TTCTCTCTAT	GGAGGAACGC	CCGAGGCGAC	CGCATCTCGA	AGATG	1945

Fig. 20

1	CTGCAGCAAC	AGATAATAAC	GAAATCAAAC	CCTCAACAAA	CCTAATAAAA	50
51	AAAATACTAA	ATGGTGCTAT	CTAATACCCA	GTTCTGGATA	TATGGAGTTG	100
101	TAGTGC GCCT	CAGCCCAAAA	ACTAATCATA	TAAGTGAAC	AGCAAGACTT	150
151	TAGATAGCAA	CACCACGAAT	CAGTCTTCTA	AAACTTCACT	AGTTTATTAT	200
201	CACCAGCGGA	TGCAAACAAA	GCTCCACTCA	CACTGAAGCA	AATAGCTCTT	250
251	ATAGCGTCTG	AATGAGAACG	ACCACCACAA	TCATCTGACA	ATGAAACTGG	300
301	ATGACCAGCC	CTGGAGGAAC	AGAGTAAATA	TTTACTTGCA	CATTCCACCA	350
351	AATGAAATGC	GGAGAAAAGG	CGCTCCACTT	CTCACTGAAA	CATGTGAACT	400
401	CAGGATTCAA	CAATATAAAA	TTAAGCAACC	AGGACCTGAT	TTCGTTAGTA	450
451	ACAATTTACA	CAACCTGACT	CACAAGAATT	CCATTGTTTT	CTTAAAGAAA	500
501	ACTTTCCATT	CCTCTCCACA	CGAAACTAAG	TTATAAGAGA	AAACAACTAA	550
551	CGATAAGCAG	CAGATAGATT	CAGTTCAGAA	ACGCACTACA	ACCTCCACCA	600
601	AAATCTACCA	ATCGTGTAAT	CAAACACTAG	CACTTTCTGC	ATAAGAGGTC	650
651	CATGTTTAAT	TAGGAAGCGG	TGATTGAAGC	GGGTATTCCA	ATTCGAAAC	700
701	CTGAGATCGA	ACACGCGGAG	CTCGGGCCCA	ATGGCGACGG	CGACGGAGTT	750
751	GCCGTGCGGG	TGGGCGCGAC	GAGCGCCGGC	GCGAACTCCG	CCGCGCCGCT	800
801	TACCCTCGGG	CTCTTCGATC	GCTGTGTCTT	CCATTGCAGC	GGCAGCCGCA	850
851	AGATAGGGAG	CTCGGATTCT	AATCGAGCGC	GAGACGGAGG	CGGGGAGGGA	900
901	TTCAGGGATT	AGGGTTTACA	GGCAGTCGCA	CATGGGGCCC	AGAGACCACT	950
951	GTCACGAAGG	AGAGCCCAGG	TGGGTTGCAG	TTGCAGGCTT	GCAGCCCAAA	1000
1001	GGGCAAAAGC	CTTTGGTTTG	TCGGTCATGG	GCCTCCACGA	AAACTGTCTC	1050
1051	TGTCGGACTG	CCCAATCCCA	CGGAAGGCCG	AGATGAACGC	AGCACCTCGA	1100
1101	TGAACGCCTC	AGATTCGCCA	ACCCACACGA	CCGCACCTAT	ATAAAGTATC	1150
1151	CACCTCCGC	TCCCGTCTTC	CTCTCCGCCG	CCGCCACGCG	CTCGCCTTTG	1200
1201	CTCCCTCTGT	TTCCCTTCAT	AGAAAGGCCG	AGGCCAGGAA	CGCCCAAGGC	1250
1251	GACCGCATCT	CCAAGATG	1268			

Fig. 21

1	CTGCAGCTAG CTGATGATCG AGACTCCGTT TGAAC TTTGA AATGTACGAA	50
51	TTTTCTTGAA CCATGTGAAA TCGGAGTCAT TTTGCACAAA ACTTGTGAAT	100
101	GAAGATCCAA ATATTCCTAA CGTATAGAAC TTCAAAGAAA GCTGGAGAAA	150
151	ATGAAAGAAA GATCACTTAT AGAAAAAAA AATGAAACTG AACTTTGCCT	200
201	TGCCACCGAT GACATCAAAA GCATTTTCGC CTAATGCTTG TAGTGGACCG	250
251	GCCTTGAGCT AGCTGCCATG TGATCGCTGA TCCTTTCGGC AGTGATGAGC	300
301	TAGCTATGCT ACTTCACTGA AGCGATGATG AGGTGTCTCG CTGCCCCCAT	350
351	TGCGGTGTAC CACAAAACGA GTCTGACCTC GCTGTCCTGC TGATGGCATC	400
401	CTTACTTGCT TTTTCCATTA TTTTGCAAGG CAAAGTTGAT CCATGGACAA	450
451	CTACTCCCGC AGAACAGTTC AATGGGCTCA AATATTTCTA TGCTAGCTTT	500
501	TTGATAAAG GTGGTGGTGC CTAATGTTGT CTAAAGCAAG GAGACGGACT	550
551	TGACCCAAAG TTGATAAGGG TCTCATCCAT TTGCCTTGAT TAAGCGGAAC	600
601	AAGACACTTG ATAGGAATAG GTTTGGTTTT TACCTAAAAT GATGCACATG	650
651	AAAGTTATTT TTTGTCAAAC TCCAAATCCT CAAATAGCTT ACCAAAGTTT	700
701	TGGCCAAATT TAGATTTGAA AATAAAGTAT AGTGTGCAAT AATTATGTTG	750
751	TCTACCTAAA CTTTTTTTCC ATCAAATAAA AGTTCAGAGT TTTTAGTGGG	800
801	TGGTGATTGT TATATAGGGG GTCGACACGG AGCTCTTTTA ATGAACTAAT	850
851	CTAAGTTTTC TAATAATCTA TATCTAATAT CTGTCATCCT TTGTCCCTTA	900
901	CAACTGTCAG ATGGAGATTG GACGAACTCA ATCCCTTCAA TTCTTATACC	950
951	CATACAAGCT AGAGCGACAC GCATCTGGGG CACACTGTGG TGTTGATTT	1000
1001	GCAAACGAAT TGAAACGCAT GATGACATGA TCGCTAAATA AATCTCCAAG	1050
1051	CCGCAAGTCT TCTAGCAAGT AACCACGCAA GAAGTTAATT GTCTTATTGC	1100
1101	AGCGCACGGG TATATTTGCT AGTTATATTA ATAAGAGAAA ATTTCTTCAT	1150
1151	CCAAATTTTT GTTCGTCCCT CTCTCCCGAT CCATGTAACGT GTCAACTCCC	1200
1201	TTGAGGGCCA ACAATAAGAG AATAGTGGTA CGTGATGAAT TAAATATAAG	1250
1251	GGTTCTAATA GGTTTACTGT TTTGTGTTGT GTCAAACCTCA CCGCCGCGCC	1300
1301	CATATACGAT TCAACTAATC TTTGGATTAC CGTAACTTGA CCTGACTGTA	1350
1351	TCAAAACTCC TTTTATTCTG CTTAATGAAA TACGTGCTAA AGCAGCATCT	1400
1401	CGAAAAAAA CTACAGCACA GTGTCCAATT TCAAGATATA TTAGAGCAAA	1450

Fig.21 cont'd

1451	TGATAGAAAT	TGTATATCCT	GTACATTGCC	GCACACGAAA	ATATTTGCTA	1500
1501	ATAATAAAAA	AGATGCCATA	AAATTGCTTG	AAGCTCCTGG	TAATAAGCAG	1550
1551	CTGGTAAATA	ATTCTTAA	ACGAGAAAAG	AAGAACCTCA	CTAATATCTC	1600
1601	CTTTGCTTAC	CTTCAGTTCA	TCAGCCAACG	ACGAGGTAGG	GTTCAGTATC	1650
1651	ATGATCCAAT	TATCCCATCG	TGACATAGCC	TTGTCCTTGA	TGATTGAGA	1700
1701	TGCAATTCTA	ATTCTCATCA	TATCATCGAC	TAGGTAACAC	AGAAAACAAA	1750
1751	CTTTTTTCTT	CGTCAATTGC	ACTGCAACCG	TTGCTTTTTG	TGATGTGCAG	1800
1801	TTGTGCACCC	ACATCACAAC	AGCACTCAAC	ACATGCCACT	TCAAAGTTCG	1850
1851	AATCGACACC	AGAACTGACG	GGAGAAAAGA	AAACAAATTA	ACAAAACGTG	1900
1901	AGAATAGATC	ATCCAGTCAT	CCAGCGTCCA	AAAAGTCCTG	CTAGCTATAG	1950
1951	ATGCAACCTA	ATAACTTTGC	TGACCTAGTC	ACTCCGAATT	CCAACATCAC	2000
2001	ATCATCGTAG	TAGGCTCATT	GTCATAGCAT	TCCTCAACAG	CACTGTTAAC	2050
2051	AAGCAGATGC	AAACAAGCAG	ATGCACATTC	ACCGGTCCCA	ATTGCACTGT	2100
2101	TAACCCGACA	ACCGCCTCTA	TTCTGTGAC	CTCTAATCAC	GTACGAGCAA	2150
2151	AGCACTAATC	AATCTCTCTC	CCCTCCCTG	TAACCAAAGC	TGCCGAATCT	2200
2201	CTTCGCTGAT	CTGGCTGCTG	CACCGCTGCA	GCCTGCGAGG	AGAGGTGGGG	2250
2251	GTCGCTCATC	AGTTAACTAC	TGACGCCAAG	GACGGCGTGC	AACGTGCAGC	2300
2301	AACAATGAGG	CGCAGGATGC	ACACCTCACA	ATGCCCTGCA	CCGCAATGAA	2350
2351	GCCTACATCT	CCCGAAGGTA	CACCTGTCTC	CATGGACACA	CATGCTGGAA	2400
2401	CCTGGAGAGA	TGCAATGCAAG	CAAGCAGACC	ACACATCATC	GATCGGTCTG	2450
2451	TGAACCTGTG	ATCACTTGGT	GTTGGTGATC	TATCGATCTC	TCCCACAGAT	2500
2501	TCACGCACAG	GGGCCTGCTG	GGTGAAAGAA	CCAAGACACC	GTGCACACCG	2550
2551	CCCCTTTTGG	ACCCTTTCCA	TGTGTCATGA	CACGGTCATG	GGCACCTTTT	2600
2601	GGTTGCATTG	CATGACATGT	TCATGTGTGC	TCGTACAGCT	CCTCAAGATT	2650
2651	CCCGATCATG	ATACCAGCGA	CGACAGCTCT	TTATTAGATC	AAGGGGGATT	2700
2701	TTTAAAAAAA	AAATCGGCAG	AGTGCTAAGA	AGCCCTTTTG	TGTTCTCCCC	2750
2751	ATTCTGGCCT	CGCCGCCGGG	CCTCTATATA	TAGCGCTCCC	ATCTCACCAC	2800
2801	CTTGCTTCAC	ACAAGCTCGA	ATGCTCGATC	GATCCCACTC	TCAGCTGATC	2850
2851	GCTCACTCTT	GCAGCTCGAT	CAGTCTTAGC	TCTAGCCTCT	AGCTAGCCAA	2900
2901	CTAGCCACTC	CTTCGTGTAG	CCATCAGCET	TCTGATCGTC	ACCAATG	2948

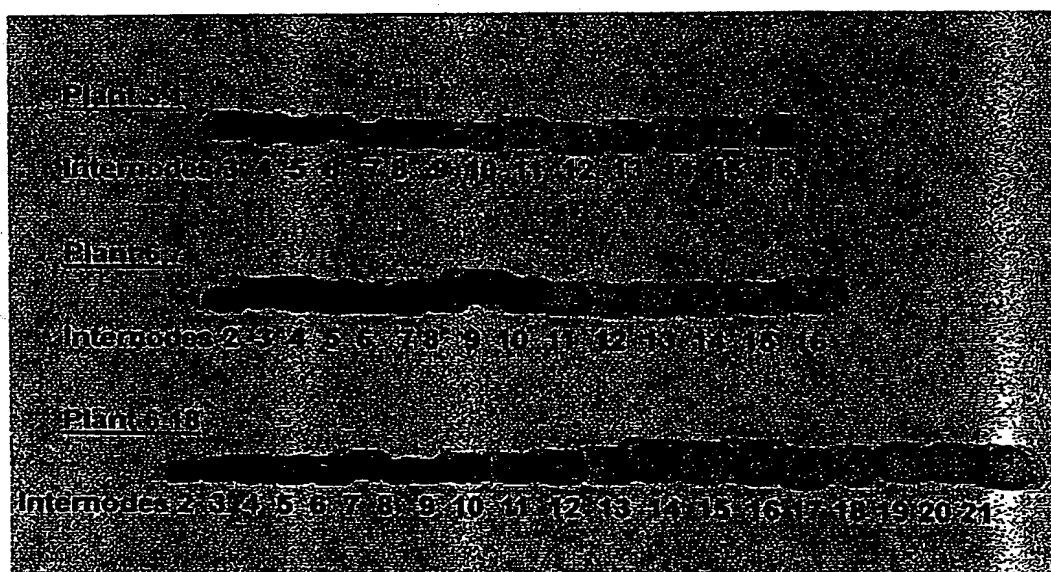
Fig. 22

Fig. 23

1	TCCTTTATTT	CATTCATTCG	TTCATCCGTG	TGAATTGGTA	CAGGTGCATA	50
51	AAGAAAAGCT	TAGCTAGCCC	GCCATTTTAA	GGCAGCCGGG	CAGGTATACG	100
101	GTGTTGACAT	ACATACATAC	AAGTAGGACG	GATTTACAAC	GCTAGTTGTG	150
151	GCAGATTTGA	ATAAAAAGAA	GTAGCAAATT	ACACGCGAAT	TCGAATCCAT	200
201	TCAGTTGTCG	AGGTGTACTA	TATAGCAAGC	TCATCGGAAG	ACGACGACGA	250
251	CGTACGGTTC	GTTCGTTCGT	TTATTGACAA	ATAATTCCAA	ACTTGCACCT	300
301	GGATGCATGA	TGATATACGT	ACACGTGTAC	GTACTAGCAG	GTTGGATCGG	350
351	ATTGATCACC	AGGAGA	366			

Fig. 24.

1	TTGAGTACTG AGTAGCTAGC TAGCCCAACT GTTTACTTGC TTTCCCTTTC	50
51	GCGAGCTGGG TGGGGTGGTT TGCGCTTCAA AGGCCGCGGC GCTTCTCTGG	100
101	CGGCCTCACG CCGGAGGCCT TGCCGTCGCT GAAGAAGCTT TTGTCGGGGA	150
	S G K	
151	AGCTGGAGAA CAACCAGATC CCTTGGCGTG GGGACTCAGC GCTGACCGAT	200
	L E N N Q I P W R G D S A L T D	
201	GGGAAGGAAG CGGGACTGGA TCTTTCCAAG GGAATGTATG ATGCTGGGGA	250
	G K E A G L D L S K G M Y D A G D	
251	TCATATGAAG TTCACCTTCC CGATGGCATT CACGGCGACG GTGCTCGCGT	300
	H M K F T F P M A F T A T V L A W	
301	GGTCGGTGCT GGAGTATGGG GATCAGATGA GCGCGGCAAA GCAACTGGAC	350
	S V L E Y G D Q M S A A K Q L D	
351	CCTGCCCTCG ATGCACTGAA GTGGATTACT GATTCCTTAT CGCTG	395
	P A L D A L K W I T D S L S L	

Fig. 25

INc IN Nc N R YL ML OL M



Fig.26

```
1  ATGTCGTGCT GCGGAGGCAA CTGTGGGTGC GGCTCCGGCT GCAAGTGC GG 50
   M S C C G G N C G C G S G C K C G
51  CAGCGGCTGC GGAGGGTGCA AGATGTACCC GGACATGGCT GAGCAGGTGA 100
   S G C G G C K M Y P D M A E Q V T
101 CCACCACCAC CCAGACTCTC ATCATGGGTG TTGCACCATC CAAGGGGCAC 150
   T T T Q T L I M G V A P S K G H
151 GCCGAGGGCG GGTTCGAGGC GCGGCCGCC GGAGCTGAGA ACGACGGCTG 200
   A E G G F E A A A A G A E N D G C
201 CAAGTGTGGC CCCAACTGCA GCTGCAACCC CTGCACCTGC AAGTGAGCTG 250
   K C G P N C S C N P C T C K
251 ATGACCCTGC CGTGATGAGG AGATCGGAAG GACTCTAGCT AAGCTCTATT 300
301 ATAGCTGTGT CGTTTGTGTC TCAGTCAGTC AGTCGCGTGC CATGGCATGC 350
351 GTAGTTGCTC GCTCGCATGC CGCTAGTGGT GGTATTGATG GGAACCAATA 400
401 ATTGTACTGA GCCGTCGAGT CCTGTCCTTC CTTCCCCTGC CCCTGGGGCT 450
451 CATCTAGCAG TCGGTGTTGT GTTTGTGGGT GCAGCACGGC AGCAGGCCAT 500
501 GCCCCATGCT CTGTCTGCAC GTCTTGTCGG GCTTTGTTTG TGCTGTGTTA 550
551 CCGTCCATCT ATATCTATAT CCCATCTTTG TGAACAAAAA AAAAAA 597
```


Fig. 27

Pindar	NCo310
L S R	L S R



Q110	Q145
L S R	L S R



Fig. 28

IN₁ IN₄ IN₆ IN₈ IN₁₁ IN₁₅

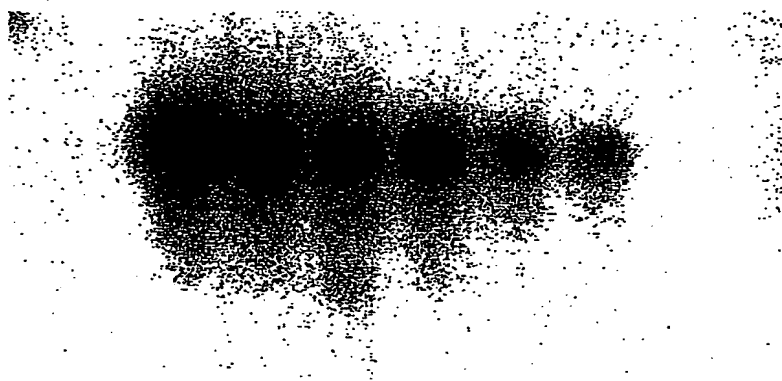


Fig. 29

1 CCACACGAGG ATCATTCACT CACTACAGTG GCGTACTAGA ATACCACTAG 50
51 TCGCGACACA CGAGCAGCAG CCGCCGTGCA TAGAGTAGTT GTGATGTACA 100
101 GGTAGTAGCA GCAGCTCGGC TCCATGGAGG ATCTGTACAG CATCCACCCG 150
M E D L Y S I H P
151 GGGATCTCGC GGGTCGGCGG CGCGGCGAGC GAGGCGTCCA CCGCCGGCGT 200
G I S R V G G A A S E A S T A G V
201 CGGCGCGGGC GGCCCTCGC CGTCTGATCT GACGGAGCTC ATGAAGGCGC 250
G A G G P S P S D L T E L M K A Q
251 AGATCGCCAG CCACCCTCGC TACCCCTCCC TCCTCTCCGC CTACATCGAG 300
I A S H P R Y P S L L S A Y I E
301 TGCCGCAAGG TGGGAGCGCC TCCGCAGGTG GCGTCGCTAC TGGAGGAGGT 350
C R K V G A P P Q V A S L L E E V
351 CAGCCGGGAG AGGAGCCCCG GTGCCGCCGG CGCCGGGGAG ATCGGCGTCG 400
S R E R S P G A A G A G E I G V D
401 ATCCCAGAGT CGACGAGTTC ATGGACTCTT ACTGCCGGGT GCTGGTGC GC 450
P E L D E F M D S Y C R V L V R
451 TACAAGGAGG AGCTGTGCGG GCCGTTTCGAC GAGGCCGCGT CGTTCCTGAG 500
Y K E E L S R P F D E A A S F L S
501 CAGCATCCAG GCGCAGCTCA GCAACCTCTG CAGCGCCGGC AGCTCGCCGG 550
S I Q A Q L S N L C S A G S S P A
551 CGGCGACCGC CACGCACTCC GATGACATGA TGGGGTCGTC TGAGGATGAG 600
A T A T H S D D M M G S S E D E
601 CAATGCTCAG GGGACACTGA TGTGCCAGAC ATAGGGCAAG AACATAGCTC 650
Q C S G D T D V P D I G Q E H S S
651 TCGCTTAGCT GACCACGAAC TCAAGGAAAT GCTTCTTAAG AAGTACAGTG 700
R L A D H E L K E M L L K K Y S G
701 GATGCCTCAG CCGTCTTCGT TCGGAGTTCC TGAAGAAGCG GAAGAAAGGG 750
C L S R L R S E F L K K R K K G
751 AAGCTACCGA AGGATGCACG GACAGTATTA CTAGAGTGGT GGAACACGCA 800
K L P K D A R T V L L E W W N T H
801 CTACCGCTGG CCTTATCCTA CGGAGGAAGA TAAGGTGAGG CTTGCAGCGA 850
Y R W P Y P T E E D K V R L A A M
851 TGACCGGTCT CGACCCAAAG CAGATCAACA ATTGGTTCAT CAATCAGAAG 900

Fig.29 cont'd

T G L D P K Q I N N W F I N Q K
901 AAGAAGCATT GGAAGCCATC CGAAGACATG CGGTTCCGCGC TCATGGAGGG 950
K K H W K P S E D M R F A L M E G
951 TGTGCGCGGT GGATCTTCTG GGACGACATC TACTTCGATA CAGGCACAAT 1000
V A G G S S G T T S T S I Q A Q L
1001 TGGACCCTGA ATCACACACC APTTGGGATG ACAATTGGCC AATTCAATCA 1050
D P E S H T I W D D N W P I Q S
1051 GTAGTAAGAC CTGGCATGTG AAGTGACGAT CTGCCCCGGTC AAAATTGACA 1100
V V R P G M
1101 ATAAATCTGT CGAGCTGAGG TTGATCACAT TAGTCAGTTG CCCCAGATCA 1150
1151 TGTGTATATG GTGCCATCGT ATCAAAACAA ACTGTATGTA TGGGCGAATT 1200
1201 GAGGAGACCT GCAAAAGCAT TTAATTAGTA GTTTCACGTA TTGGCTCATG 1250
1251 GATTTGTAAT ACTCGCTACC CAATTTAATT TTTAGATAGG CTGAAGGGCT 1300
1301 TATAAAGATA AAATTACTTC TGCAAAAAAA AAAAAAAA 1339

Fig. 30

Pindar	NCo310
L S R	L S R



L S R	L S R
Q110	Q145

Fig. 31

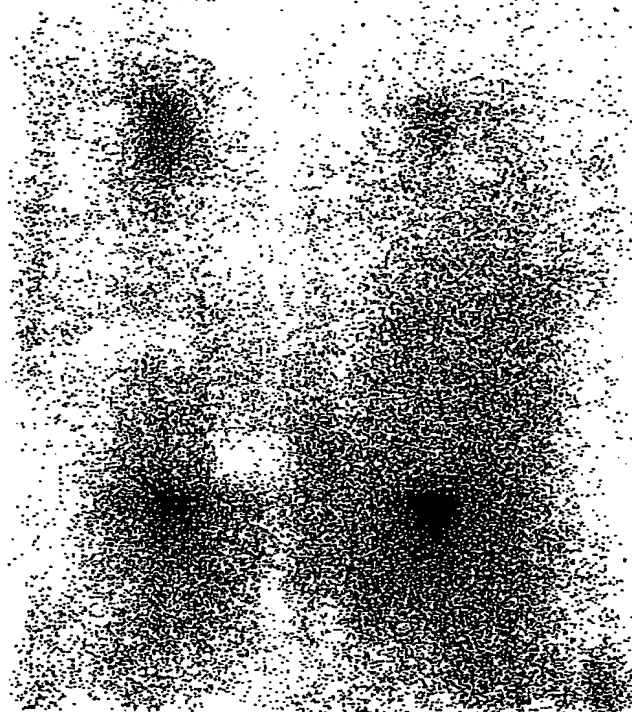
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1  TGAGCAGCGT CGTCACCGCC GCCGCCGCCA TTCAGGCTGA CCCC GCCGAG 50
   S S V V T A A A A I Q A D P A E
51 AAGCCGCGCG CCCCCAAAGC TCAGGTGGTG GGATGGCCAC CGGTCCGGTC 100
   K P R A P K A Q V V G W P P V R S
101 GTTCCGGAAG AACATCATGT CGGTGCAGTC CGACAAGGGC GCCGGCGGCA 150
   F R K N I M S V Q S D K G A G G S
151 GCAAAGACGC CGACAAGTCC TCGCCGCCGC CGGCGGCAGC AGCAGCAGTT 200
   K D A D K S S P P P A A A A A V
201 GCGGTGCGG CGTTCGTGAA GGTGAGCTTA GACGGCGCGC CATACTGCG 250
   G G A A F V K V S L D G A P Y L R
251 CAAGGTGGAC CTCAAGATGT ACAAGATGTA CAAGAGCTAC CAGGAGCTGT 300
   K V D L K M Y K M Y K S Y Q E L S
301 CCAAGGCGCT CGAGAAGATG TTCAGCTCCA CCATTGGAAG CTGTGGGTCT 350
   K A L E K M F S S T I G S C G S
351 CAAGGGATGA ACGGCATGAA CGAGAGCAAG CTGGTGGATC TGCTCAACGG 400
   Q G M N G M N E S K L V D L L N G
401 CTCCGAGTAC GTGCCGACCT ACGAGGACAA GGAGGGCGAC TGGATGCTCG 450
   S E Y V P T Y E D K E G D W M L V
451 TCGGCGACGT GCCGTGGGAG ATGTTCTGTCG AATCATGCAA GCGCCTTCGG 500
   G D V P W E M F V E S C K R L R
501 ATCATGAAAG GATCAGAAGC CATTGGCCTC GCGCCGAGGG CCATGGAGAA 550
   I M K G S E A I G L A P R A M E K
551 ATGCAAGAAC AGAAGCTGAG GAGAAGATGG AAGCTTGCAT CTGAACACCA 600
   C K N R S
601 AGGATGGATG CAAGAAGATC CATCTCCACA GCTTTTGCAG CTCAGCTCAT 650
651 CTCTGTATCA TACTCGTCTC TGTCCTCCAA GCCTCTATAT CTCCTGATCA 700
701 ATGAAGTCAT TTTGGTGTGT CAACTGTTCT ATCTGTGCTA GTTTGTGAGT 750
751 AAACGAGTCA GTCAGTGTTT AAGTCTCATC TTGTCCTATA CAGAAGAGAG 800
801 AAGACATGAG CATGTGTACC TGTCCTATAA CAATGTCTTT TTGCACTACC 850
851 AGTTCTCTCC TCTTCTTCTT TTTTCTTTT TTTTGGTGG CACAAAATGC 900
901 AAAGGCACCC TGCTCCTAAT CAAGGGGCCC TTTTGCATCT TGTCCATCAA 950
951 AAGTTGTTTA CTTATAATTA TCACGGTGTG TCAGTGGAAC AAATAATTGT 1000
1001 TGCAAGTTGT TAAAAAAAAG CTGTATTGTA TGAAAAAAA AAAAAA 1046

```

Fig. 32

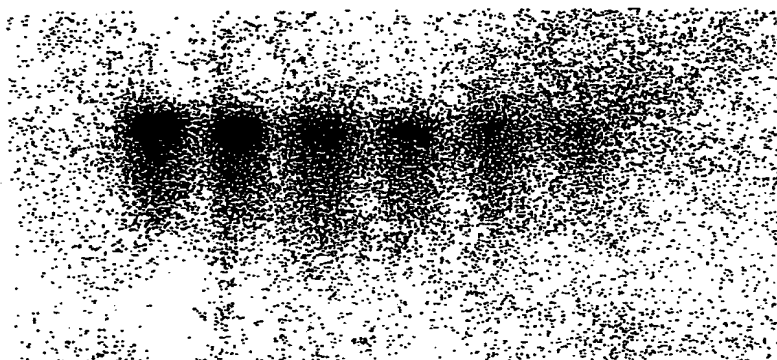
Pindar **NCo310**
L S R **L S R**



L S R **L S R**
Q110 **Q145**

Fig. 33

IN₁ IN₄ IN₆ IN₈ IN₁₁ IN₁₅



INTERNATIONAL SEARCH REPORT

International application No.
PCT/AU00/01033

A. CLASSIFICATION OF SUBJECT MATTER

Int. Cl. ⁷: C12N 15/29, C07K 14/415, A01H 5/00

According to International Patent Classification (IPC) or to both national classification and IPC

B. FIELDS SEARCHED

Minimum documentation searched (classification system followed by classification symbols)
AS BELOW.

Documentation searched other than minimum documentation to the extent that such documents are included in the fields searched
AS BELOW

Electronic data base consulted during the international search (name of data base and, where practicable, search terms used)
EMBL GenBank : Seq ID.Nos. 7, 12, 13, 14, 15, 16, 17, 18, 19, 20, 21, 22, 23
DGENE : Seq ID Nos. 16, 17, 18, 19, 20, 21, 22, 23

C. DOCUMENTS CONSIDERED TO BE RELEVANT

Category*	Citation of document, with indication, where appropriate, of the relevant passages	Relevant to claim No.
X -	EMBL, Accession No. AF111029, A novel maize gene homologous to 40S ribosomal protein S27., Stukart G. C. <i>et. al.</i> 21 December 1998 ✓	38,39,40
X -	EMBL, Accession No. AF083336, Involvement of <i>Arabidopsis thaliana</i> ribosomal S27 in degradation triggered by genotoxic stress., Revenkova E., <i>et. al.</i> 29 January 1999	38,39,40
X -	GenBank, Accession No. HVMNE1MR, Zinc-finger protein., Huang, C. <i>et. al.</i> 16 March 1995	38,39,40

☒ Further documents are listed in the continuation of Box C ☒ See patent family annex

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Date of the actual completion of the international search
25 October 2000

Date of mailing of the international search report
- 6 NOV 2000

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INTERNATIONAL SEARCH REPORT

International application No.

PCT/AU00/01033

C (Continuation). DOCUMENTS CONSIDERED TO BE RELEVANT		
Category*	Citation of document, with indication, where appropriate, of the relevant passages	Relevant to claim No.
X	EMBL, Accession No. AC006434, Genomic sequence for <i>Arabidopsis thaliana</i> BAC F10A5, complete sequence., Cheuk R., <i>et. al.</i> , 25 January 1999	38,39,40
X	GenBank, Accession No. AC007797, <i>Arabidopsis thaliana</i> chromosome I BAC F6F9 genomic sequence, complete sequence., Nguyen M., <i>et. al.</i> 15 June 1999	38,39,40
X	DGENE Accession number 1999P-Y11496 and AU, A, 85554/98 (GENSET) 11 February 1999. See specification and sequence listing.	38,39
X	AU, B, 13087/97 (725340) (GIST-BROCADES B.V.) 3 July 1997 See SEQ. ID No. 28.	38,39
X	US, A, 5 243 041 (JOSE A. FERNANDEZ-POL) 7 September 1993 See SEQ ID Nos. 1 and 2.	38,39

INTERNATIONAL SEARCH REPORT
Information on patent family members

International application No.
PCT/AU00/01033

This Annex lists the known "A" publication level patent family members relating to the patent documents cited in the above-mentioned international search report. The Australian Patent Office is in no way liable for these particulars which are merely given for the purpose of information.

Patent Document Cited in Search Report		Patent Family Member			
AU	85554/98	EP	1000149	WO	9906551
AU	13087/97	WO	9723633	EP	870042
		EP	780474	CA	2241267
US	5243041	NONE			
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